Assiut University website: <u>www.aun.edu.eg</u>

THE INCIDENCE OF SALMONELLA ARIZONAE IN DIFFERENT POULTRY SPECIES IN NEW VALLEY GOVERNORATE, ITS SENSITIVITY AND ITS PATHOGENICITY IN TURKEY POULTS

HISHAM M.YAHI¹; MOSTAFA A. SHEHATA²; MOEMEN A. MOHAMED²; AL SHIMAA R. SAYED³; MARWA M. SAFWAT² AND AHMED K. HASSAN²

¹Department of Poultry Diseases, Agriculture Research Center, Animal Health Research Institute, New Valley Lab, Egypt

² Department of Avian and Rabbit Medicine, Faculty of Veterinary Medicine, Assiut University. Assiut.

³ Department of Poultry Diseases, Agriculture Research Center, Animal Health Research Institute, Assiut Lab, Egypt.

Received: 12 August 2023; Accepted: 18 September 2023

ABSTRACT

Arizonosis is a septicemic disease of turkey poults; one of the most frequently identified salmonella serotypes with significant mortality and morbidity causing serious outbreaks. The present work aimed to detect the incidence of Salmonella arizonae in different bird species in New Valley Governorate, its sensitivity and/or resistance to antibacterial drugs, and its pathogenicity for turkey poults. A total of 250 suspected samples from dead and sick birds of different species in the New Valley Governorate were collected. Isolation was carried out following standard methods, primarily through standard laboratory culture media, followed by biochemical identification. All obtained isolates were subjected to a set of 6 antibacterial agents to study their antibiogram sensitivity using the disk diffusion method. Experimental infection of seven-day-old turkey poults with the isolated Salmonella arizonae was carried out by oral and intraocular routes. Results revealed that 23 isolates were identified as Salmonella arizonae (9.2%) from turkey poults, pigeons, and quail of different ages. High sensitivity of Salmonella arizonae isolates was recorded for gentamicin, followed by colistin sulphate, doxycycline, and Trimethoprim/Sulfamethoxazole. High resistance of Salmonella arizonae isolates was recorded for florfenicol and amoxicillin. Clinical signs of experimentally infected turkey poults include depression, off food, whitish and greenish diarrhea, emaciation, ruffled feathers, and reluctance to move. Post-mortem examination showed that mild arthritis, bronzy coloration of the liver, typhlitis, pneumonia, hemorrhage in the brain, severe enteritis, and enlargement of spleen and gallbladder. Mortality rates in experimentally infected turkey poults reached 80% and 50% with both ocular and oral routes, respectively. We concluded that Salmonella arizonae could be detected with an incidence rate 9.2% from turkey, pigeons, and quail in New valley Governorate. Salmonella arizonae is sensitive for gentamicin, colistin sulphate, doxycycline, and Trimethoprim/ Sulfamethoxazole, while it is resistant to florfenicol and amoxicillin. Experimental infection of Salmonella arizonae in turkey poults results in high mortality and sever clinical signs and postmortem lesions.

Keywords: Salmonella arizonae, incidence, pathogenicity, sensitivity, resistance.

Corresponding author: HISHAM M.YAHI

E-mail address: hishamyahiayahia@gmail.com

Present address: Department of Poultry Diseases, Agriculture Research Center, Animal Health Research Institute, New Valley Lab, Egypt

INTRODUCTION

A diverse number of serovars of *Salmonella*, including *Salmonella arizonae* (*S. arizonae*), *have been* registered in all countries of the world. Despite the rapidly growing poultry industry, the periodic outbreaks due to *S.arizonae* infections led to huge problems in production and egg hatchability (Jones *et al.*, 2002).

Poultry species were important reservoirs of many zoonotic pathogens, of which *S.arizonae* is of prime importance to human by consumption of contaminated poultry meat and eggs. It causes gastroenteritis in young infants and immune-compromised individuals. Furthermore, the invasive form leads to complications such as meningitis, septicemia, and osteomyelitis (Cristina, 2017).

The genus Salmonella consists of only two species, Salmonella bongori and Salmonella enterica, with the latter divided into six subspecies; S. enteric subsp. enterica, S. enteric subsp. salamae, S. enteric subsp. arizonae, S. enteric subsp. diarizonae, S. enteric subsp. houtenae, and S. enteric subsp. Indica (More et al., 2017). S. arizonae is a Gram-negative, non-spore-forming, motile, rod-shaped, facultative anaerobic bacterium (Wang et al., 2015). S. arizonae infection is quite common among different poultry species including turkeys, chickens, ducks, quail, and pigeons (Özkalp, 2012). Turkeys were the principal species, especially turkey poults for infection with S. arizonae (Hafez, 2013; Tracy et al., 2020). S. arizona is transmitted through vertical and horizontal transmission (More et al., 2017).

In acute *S. arizonae* infection, young birds showed septicemia, neurologic signs, and mortality with a duration typically of 3-5 weeks, while adult turkeys exhibit asymptomatic chronically intestinal carriage and fecal shedding for extended periods (Shivaprasad, 2013). Infection of turkeys with *S. arizonae* seriously impairs fertility, hatchability, and egg production (Ibrahim *et al.*, 2013).

In the past few decades, the emergency use of antibiotics for the prevention and control of different species of bacteria in poultry industry has been on the rise (Davies, 2010). Many antibiotics currently have a broad spectrum of activity and high efficacy in a wide range of Gram-negative bacteria infections, such as aminoglycosides, chloramphenicol tetracycline, and (Shivaprasad and Barrow, 2008). Antibacterial susceptibilities of S. arizonae are not constant, but vary from time to time, with different environments, antimicrobial susceptibility testing has been conducted worldwide (Mir et al., 2015).

There was little information on the incidence and antibiotic susceptibilities of *S. arizonae* in Egypt. Thus, our study aimed to isolate and identify *S. arizonae* from different poultry species, including turkeys, chickens, ducks, quail, and pigeons in New Valley Governorate, to detect its sensitivity and/or resistance pattern against antibacterial agents, and to study its pathogenesis in turkey poults.

MATERIALS AND METHODS

1. Specimens collection:

A total of 250 suspected samples were collected from poultry farms and back yard birds in New Valley Governorate, including 84 intestines, 83 livers, 24 yolk sacs, 9 ovaries and 50 cloacal swabs; they collected from 91 turkeys aged from one day up to 7 months old, 79 chickens aged from one day up to 6 months, 30 ducks aged from one day up to 15 days old, 25 pigeons older 6 months age, and 25 quail older 6 months age; as shown in **Tab(1).**

2. Isolation and identification of *Salmonella arizonae*:

Samples were cultured in brain heart infusion (BHI) broth (Oxoid, UK) to encourage the growth of *S* .*arizonae* and incubated at 37° C for 24 hours, then sub-cultured on bismuth

Assiut Veterinary Medical Journal

Assiut Vet. Med. J. Vol. 70 No. 180 January 2024, 263-273

sulphite agar (Oxoid, UK) and incubated at 37°C for 48 hours (Shivaprasad *et al.*, 2006). Individual colonies were obtained and stained with Gram stain. Suspected colonies were subjected to different biochemical tests,

including lactose and dulcitol fermentation, hydrogen sulfide production, indole, catalase, malonate, and gelatin hydrolysis (Mahajan *et al.*, 2003).

Age of Tissue samples							
species	birds	Ovary	Liver	Yolk sac	intestine	cloacal swabs	Total
turkeys	From 1 to 7 days		20	10	20		91
	Over 7 months	7	5		4	25	91
	From 1 to 7 days		20	14	20		
chickens	One month		3		5	15	79
	Over 6 months	2					
ducks	From 1 to 15 days		15		15		30
pigeons	Over 6 months		10		10	5	25
quails	Over 6 months		10		10	5	25
Total		9	83	24	84	50	250

Table 1: Types and number of samples collected from different poultry species:

3. Detection of antibacterial sensitivity and/or resistance patterns of *S.arizonae* isolates :

All S. arizonae isolates were subjected to in *vitro* antibiotic susceptibility testing against 6 antibacterial agents of different classes, including doxycycline, gentamicin, florfenicol, colistin sulphate, amoxicillin, and trimethoprim/sulfamethoxazole (Oxoid, UK) using disk diffusion test. The diameters of inhibition zones were interpreted according to the National Committee for Clinical Laboratory Standards (NCCLS sub-Committee's recommendation, 2002).

4. Experimental Infection:

Sixty-one-day old turkey poults were maintained in separate units at the Poultry Diseases Department, Animal Health Research Institute, New Valley Governorate, Egypt, with approval from the National Ethical Committee of the Faculty of Veterinary Medicine, Assiut University,

Assiut, Egypt according to the OIE Standards for use of animals in research, following (Under ARRIVE guidelines the No.06/2023/0105). They randomly divided into three equal groups; 20 poults for each group and provided with feed and water ad libitum. Turkey poults were challenged with 0.5 ml of S. arizonae containing 2 $x10^8$ colony-forming units/mL (CFU/mL), in group (1) by ocular route and in group (2) by oral route, at 7th days old of age. Turkey poults of group (3) were kept as non-infected control group. Turkey poults in all groups were observed daily for signs, mortalities, and postmortem lesions for 14 days postinfection.

RESULTS

1. Incidence of S. arizonae:

Twenty-three suspected *S. arizonae* isolates were obtained from different poultry species in the New Valley Governorate, with a incidence rate 9.2%. A high rate of *S. arizonae* incidence was reported in turkey with a percentage of 20.8% (19 out of 91 examined turkeys), especially in turkey poults aged from 1 to 15 days. Lower incidence rates of *S. arizonae* in pigeons and quails were recorded with a percentage of 8% for both, especially in adult birds older than 7 months. Negative trials were done to detect *S. arizonae* in chickens and ducks.

In the present study, the results revealed that the highest rate of *S. arizonae* isolation was from the intestines (14.3%), followed by the liver (9.6%), and yolk sac (8.3%), respectively. *S. arizonae* was isolated from cloacal swabs with incidence rate 2%. No isolation of *S. arizonae* could be obtained from ovaries, as shown in **Tab(2)**.

Table 1.	Incidence	of C		:	different	
Table 2:	Incluence	01 5.	arizonae	m	amerent	poultry species.

	No. of		Suspected S. arizonae isolates						
Species	examined samples	Liver	Yolk sac	Intestine	Ovary	cloacal swabs	of isolates	Incidence	
Turkey	91	8	2	8	0	1	19	20.8%	
Pigeon	25			2			2	8%	
Quail	25			2			2	8%	
Chicken	79			0			0	0%	
Duck	30			0			0	0%	
Total	250	8	2	12	0	1	23	9.2%	

2. Colonial, cellular, and biochemical characteristics of *S. arizonae*:

The obtained colonies of suspected *S*. *arizonae* appeared on bismuth sulphite agar as minute circular brown-black colonies with



Figure (1): *Salmonella arizonae* colonies on Bismuth sulphite agar

a metallic sheen that turned into black when time of incubation increased as shown in **Figure (1)**. Gram-stained films which made from suspected *S. arizonae* colonies revealed gram-negative bacilli.

The results of biochemical characterization revealed that suspected *S. arizonae* isolates were positive for lactose fermentation, hydrogen sulfide production, catalase, malonate, gelatin liquefaction and negative for indole and dulcitol fermentation, as shown in **Table (3) and Figure (2)**.

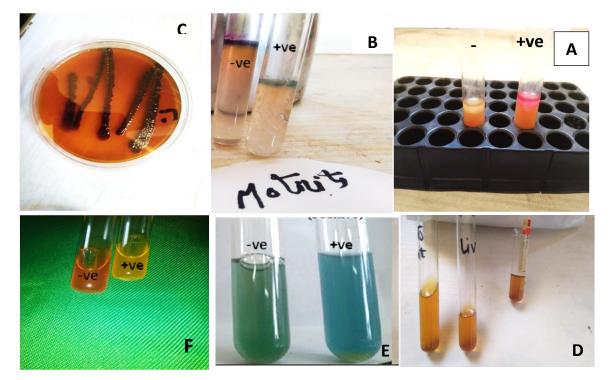


Figure (2): A: Hydrogen sulfide production, B: Motility test, C: Indole test, D: Gelatin liquefaction test, E: Sodium malonate test, F: Dulcitol hydrolysis

Table	3:	Biochemical	characters	of	<i>S</i> .
arizonae.					

Tests	S. arizonae isolates
Hydrogen sulfide production	+
Motility test	+
Lactose fermentation test	+
Indole production test	-
Dulcitol hydrolysis test	-
Gelatin hydrolysis test	+

3. Antimicrobial sensitivity:

In vitro antibiotic susceptibility assay showed that all suspected *S. arizonae* isolates were sensitive to Gentamicin (10%), Colistin (10%), Doxycycline (30%), and Trimethoprim/Sulfamethoxazole. While they were resistant to Florfenicol (10%) and Amoxicillin (25%), as shown in **Figure (3).**

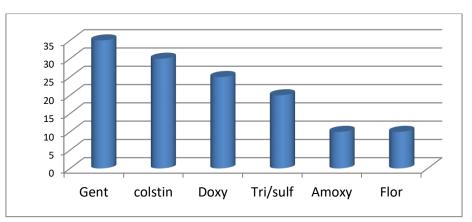


Figure (3): Antibacterial sensitivity testing of suspected *S. arizonae* isolates. Gent: gentamycin, Doxy: doxycycline, Tri/sul: trimethoprim/sulfamethoxazole, Amox: amoxicillin, and Flor: florfenicol.

Experimental infection:

In group (1), after 48 hours of ocular challenge with *S. arizonae*, *a* high mortality rate was detected (80%), and the living infected poults showed depression, offfood, whitish diarrhea, emaciation, ruffled feather, and mild arthritis. In group (2), the mortality rate reached 50% after 72 hours of oral infection and infected poults showed depression, off food, and greenish diarrhea. Bronzy discoloration of the liver, typhlitis, and pneumonia were detected in post-

mortem examination in both groups, but in group (1), there was hemorrhage in the brain, severe enteritis, and mild arthritis. Enlargement of the spleen and gallbladder was observed in PM inspection in group (2). All turkey poults in the control negative group (3), showed no clinical signs and no post-mortem lesions until the end of the experiment, results of experimental infection are shown in **Table(4) and Figure** (4,5).4.

Table 4: Clinical signs and PM lesions of S. arizonae isolates experimental infection in turkey poults.

Group	Inoculation rout	Clinical signs	PM lesions		
1	intraocular	A high rate of mortality was detected (80%) after 48 hours of infection with signs of ruffled feathers, whitish diarrhea, and emaciation.	Bronzy coloration of the liver, typhlitis, pneumonia, hemorrhages in the brain and severe enteritis, mild arthritis.		
2	orally	Low rate of mortality, mortality reached 50% after 72 hrs of infection, off food, greenish diarrhea, Emaciation.	Emaciation, the bronzy coloration of the liver, typhlitis, severe enteritis, enlarged spleen and gallbladder.		
3	Control	No mortalities and no clinical signs.	No PM lesions		

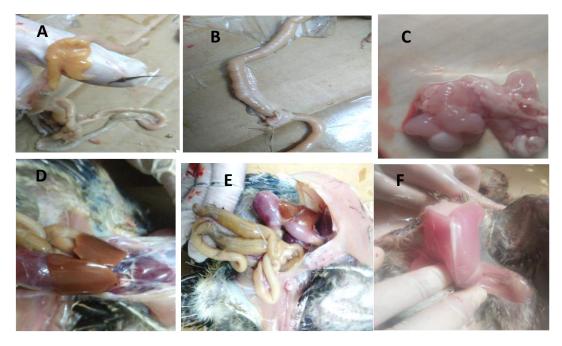


Figure 4: A,B: Severe enteritis, C: Hemorrhages in brain (ocular infection), D: Bronzy coloration of liver, E: Typhilitis (cecal core), F:Emaciation.



Figure (5): A: Sever emaciation, B: Mild arthritis (ocular infection), C: greenish diarrhea (oral infection).

DISCUSSION

Arizonosis, a disease that causes high mortality and morbidity rates in young turkey poults, resulting in severe economic losses through continuous outbreaks. Salmonella arizonae was isolated from turkey poults, pigeons, and quail in New Valley Governorate with an incidence rate of 9.2%. High rate of isolation was obtained from turkey poults (19 out of 91 samples). These results matched with a previous study, in which turkeys were the principal species for infection with S. arizonae (Park et al., 2008). More et al., (2017) also reported that S. arizonae infection was confined to the first few weeks of life in different poultry species, especially young turkeys. Carrique-Mas and Davies (2008) reported that S. arizonae infection in turkeys was around one month of age. However, they also noticed infection in laying hens with an incidence rate (6%). Our results revealed that the rate of S. arizonae isolation in pigeons and quail was (8%), this low rate of isolation may be due to the low number of quail and pigeon farms in New Valley Governorate, strict isolation and biosecurity around farms, and intermittent shedding of organisms through fecal samples,

these findings agree with Shivaprasad *et al.* (2006) and Ahmad *et al.*, (2019).

In the present work, negative trials were done to isolate *S. arizonae* from suspected cases of chicken and duck, and this was in line with Cardinale *et al.* (2005) and El-Tawab *et al.* (2015), who attributed these results to the period of sample collection, different examined organs, the hazards of antibiotics for chickens, the variable fecal excretion of the organisms, and/or the limited reliability of cloacal swab cultures, although there was another report about *S. arizonae* infection in chicken and duck in which the pathogen was completely identified and the lesions studied in some details (Izat *et al.*, 2014).

Among the examined internal organs, the highest rate of isolation was from the intestine (14.3%), followed by the liver (9.6%). These results were agreed with Shivaprasad *et al.* (2006), who indicated that *S. arizonae* could be isolated from the liver of infected poults, and with Bhunia (2007), who stated that *S. arizonae* was isolated mostly from the intestinal tract, indicating that the main reservoir of *S. arizonae* was the intestinal tract of animals and birds. Acute infections

cause septicemia, and the liver was the first affected organ.

Ibrahim et al. (2013) identified one isolate of S. arizonae from the liver of broiler chickens during their investigation of 102 samples from Omdurman farms. In another study, Lenev et al. (2016) examined 371 samples from different broiler houses in Russia, and the isolation rate of S. arizonae was 6.5% of the total isolates. The percent of S. arizonae isolation by Klishchova and Nazarenko (2021), was 6%. In our study, the lowest rate of S. arizonae isolation from tissues was the yolk sac (8.3%); this matched with Lamas et al., (2016) and Klishchova and Nazarenko (2021), who isolated S. arizonae from the yolk sac (6%); this may be explained by the severity of infection and/or antibiotics used at the point of lay, which transferred from adults to poults.

In the present study, collected samples were cultured in brain heart infusion broth to enhance the growth of S. arizonae, then subcultured on bismuth sulphite agar for differentiation of S. arizonae from other Salmonella spp. The bacterial colonies appeared minute and circular, brown black with metallic sheen because of hydrogen sulfide production by S. arizonae. The colonial and cellular morphologies and biochemical properties were similar to those reported by Murray et al. (1999). The total S. arizonae incidence rate was 9.2%, and this recent increase in its incidence among turkey poults could cause serious impediments to the poultry industry, especially in developing countries in Asia and Africa.

Our results of sensitivity and/or resistance pattern of all *S. arizonae* isolates against gentamicin and colistin sulphate were similar to those observed by Evangelopoulou *et al.* (2014), while they disagree with Irfan *et al.* (2015), who found that most of *S. arizonae* isolates were resistant to colistin sulphate; this may be due to the presence of resistance genes to colistin sulphate in some *S. arizonae* isolates. Our results of *S. arizonae* sensitivity to doxycycline was comparable to Al-Salauddin *et al.*, (2015), who reported 100% resistance to tetracycline in their study. Resistance to tetracycline has been attributed to its irrational use of it as a growth promoter in poultry feed. In recent years, the use of tetracycline has been limited in food animals, which explains the change in the pattern of resistance.

Another finding of interest is the resistance of S. arizonae isolates to florfenicol and amoxicillin. This agreed with the study by Diarraa and Malouin (2014), who reported in their study a similar pattern of resistance against amoxicillin. These findings may be due to the growing resistance towards betalactam antibiotics, that has been prevalent worldwide among members of the Enterobacteriaceae of animal origin, especially in Salmonella spp., associated with antibiotic-resistance various gene determinants. This observation draws serious attention, as poultry serves as an important source of transmission of these multidrugresistant S. arizonae genes to human.

Results of our study about sensitivity of S. arizonae to florfenicol did not match with a study by Mei et al., (2021), who used florfenicol in poultry production to prevent and treat Salmonella infection, reducing intestinal colonization and decreasing susceptibility to Salmonella infection. Routine use of florfenicol for prophylactic and therapeutic purposes in the poultry industry leads to bacterial resistance to this antibacterial agent and selective pressure on various bacteria, such as S. arizonae.

Turkey poults, the most substable species for arizonosis, were experimentally infected with *S. arizonae* isolates, leading to developing of clinical signs and PM lesions similar to those reported by Tracy *et al.* (2020).

Hemorrhages in the brain were occurred in turkey poults infected by the ocular route, which may be attributed to the vascular spread of the infection to the brain, undoubtedly following primary replication of S. arizonae outside the nervous system, which comes close to results obtained by Jortner and Larsen (1984). Also, the inflammation of the hock joints which was observed in birds that probably suffered from localization and bacteremia of the microorganisms in certain parts of the skeletal system as joints, that agrees with finding reported by Oh et al. (2010).

CONCLUSION

The present study demonstrates that S. arizonae caused serious mortality in turkey poults, as a first report in New Valley Governorate farms (turkey poults, pigeons, and quail). S. arizonae was detected in turkey poults, pigeons, and quail in the New Valley Governorate, with an incidence rate of 9.2%. The sensitivity and resistance to antibacterial agents in this study draw serious attention to the poultry sector in Egypt and New Valley Governorate, especially for the real need to prevent and control S. arizonae in poultry farms. Such data is essential for developing appropriate treatment of S. arizonae using high-efficacy antibacterial agents after conducting a sensitivity test. Proper scientific and public health regulations are needed to control the non-judicial use of antibacterial agents, that may reduce the emergence of microbial bugs, which are spreading worldwide and responsible for fatal disease outcomes in different parts of the world.

Ethical approval

The Animal Ethical Committee of the Faculty of Veterinary Medicine, Assiut University, Assiut, Egypt according to the OIE Standards for the use of animals in research following ARRIVE guidelines has approved the present study under permission No: 06/2023/0105.

REFERENCE

Ahmad, C.D.; Al-Juboory, Y.H.O. and Zenad, M.M. (2019): Isolation and diagnosis of Salmonella germs in domestic and wild pigeons. Plant Archives, 19(2), 4084-4088.

- Al-Salauddin, A.S.; Hossain, M.F.; Dutta, A., Mahmud, S.; Islam, M.S.; Saha, S. and Kabir, S.L. (2015): Isolation, identification, and antibiogram studies of Salmonella species and Escherichia coli from boiler meat in some selected areas of Bangladesh. Int. J. Basic Clin. Pharmacol, 4(5), 999-1003.
- *Bhunia, A.K. (2007):* Food borne microbial pathogens. 1st ed. New York, Springer Science and Business Media., p 276.
- Cardinale E.; Gross-Claude J.; Rivoal, K.; Rose, V.; Tall, F, Mead, G. and Salvat G. (2005): Epidemiological analysis of Salmonella enterica spp. serovar Hadar, Brancater and Enteriditis in humans and broilers chicken in Senegal using Pulsed Field Gel Electrophoresis and antibiotic susceptibility. Journal of Food Protection. 60:1312-1317.
- *Carrique-Mas, J. and Davies, RH. (2008):* Sampling and bacteriological detection of Salmonella in poultry and poultry premises: a review. Revue Scientifique et Technique de l'Office International Des Epizooties, 27, 665–677.
- Gavrilovici, C.; Pânzaru, C.V.; Cozma, S.; Mârţu, C.; Lupu, V.V.; Ignat, A. and Stârcea, M. (2017): "Message from a turtle": otitis with Salmonella arizonae in children: case report. Medicine, 96(44).
- Davies, J. (2010): Origins and evolution of antibiotic resistance. Microbiol Mol Biol Rev 2010; 74(3): 417-33.
- *Diarra, MS. and Malouin, F. (2014):* Antibiotics in Canadian poultry production and anticipated alternatives. Front Microbiol 2014; 5: 282.
- El-Tawab, A.; Ashraf, A.; El-Hofy, F.I.; Ammar, A.M.; Nasef, S.A. and Nabil, N.M. (2015): Studies on different salmonella serotypes isolated from poultry in different governorates in Egypt. Benha Veterinary Medical Journal, 28(2), 169-175.

- Evangelopoulou, G.; Kritas, S.; Govaris, A. and Burriel, A.R. (2014): Pork meat as a potential source of Salmonella enterica subsp. arizonae infection in humans. Journal of Clinical Microbiology, 52(3), 741-744.
- Hafez, HM. (2013): Salmonella infections in turkeys. In: Barrow PA and Methner U 2nd (eds.). Salmonella in Domestic Animals. CAB International, Wallingford, UK. pp. 193–220.
- Ibrahim, H.H. Emeash; Nahed H. Ghoneim and Abdel-Halim, M.A. (2013): Sero epidemiological Studies on Poultry Salmonellosis and its Public Health Importance J. World's Poult. Res. 3(1): 18-23, 2013.
- Irfan Ahmad Mir; Sudhir Kumar Kashyap and Sunil Maherchandani (2015): Isolation, serotype diversity and antibiogram of Salmonella enterica isolated from different species of poultry in India Asian Pac J Trop Biomed 2015; 5(7): 561–567.
- Izat, A.L.; Kopek, J.M. and McGinnis, J.D. (2014): Research note: incidence, number, and serotypes of Salmonella on frozen broiler chickens at retail. Poultry Science, 70(6), 1438-1440.
 - Jones, D.R.; Anderson, K.E.; Curtis, P.A. and Jones, F.T. (2002): Microbial contamination in inoculated shell eggs: effects of layer strain and hen age. Poult. Sci. 2002. 81:715–20.
- Jortner, B. and Larsen, C. (1984): Granulomatous ventriculitis of the brain in arizonosis of turkeys. Veterinary Pathology, 21(1), 114-115.
- Klishchova, Z. and Nazarenko, S. (2021): Monitoring of Salmonella infection of poultry for the period from 2016 to 2020. EUREKA: Health Sciences, (2), 97-101.
- Lamas, A.; Fernandez-No, IC.; Miranda, JM.; V_azquez B.; Cepeda, A. and Franco CM. (2016): Prevalence, molecular characterization and antimicrobial resistance of Salmonella

serovars isolated from northwestern Spanish broiler flocks (2011-2015). Poultry Science, 95, 2097–2105.

- Lenev, S.V.; Laishevtcev, A.I. and Pimenov, N.V. (2016): Improvement of allocation and identification of salmonella enterica bacteria of arizonae subspecies. Russian Journal of Socio-Economic Agricultural and Sciences, 50(2), 14-23.
- Mahajan, R.K.; Khan, S.A.; Chandel, D.S.; Kumar, N.; Hans, C. and Chaudhry, R. (2003): Fatal case of Salmonella enterica subsp. arizonae gastroenteritis in an infant with microcephaly. Journal of Clinical Microbiology, 41(12), 5830-5832.
- Mei, X.; Ma, B.; Zhai, X.; Zhang, A.; Lei, C.; Zuo, L. and Wang, H. (2021): Florfenicol enhances colonization of a Salmonella enterica Serovar Enteritidis floR mutant with major alterations to the intestinal microbiota and metabolome in neonatal chickens. Applied and Environmental Microbiology, 87(24), e01681-21.
- Mir, I.A.; Kashyap, S.K. and Maherchandani, S.J. (2015): Isolation, serotype diversity and antibiogram of Salmonella enterica isolated from different species of poultry in India. 5(7), 561-567.
- More, S.; Bøtner, A.; Butterworth, A.; Calistri, P.; Depner, K.; Edwards, and Bicout, D. (2017): Assessment of listing and categorisation of animal diseases within the framework of the Animal Health Law (Regulation (EU) No 2016/429): Salmonella infection in poultry with serotypes of animal health relevance (S. Pullorum, S. Gallinarum and S. arizonae). EFSA Journal, 15(8).
- Murray, PR.; Baron, EJ.; Pfaller, MA.; Tenover, FC. and Yolken, RH. (1999): Manual of clinical microbiology. 7th ed. Washington D.C.: ASM, 1999.
- *Oh, JY.; Kang, MS.; An, BK.; Song, EA.; Kwon, JH. and Kwon, YK. (2010):* Occurrence of purulent arthritis broilers vertically infected with Salmonella

enterica serovar Enteritidis in Korea. Poult Sci. 2010 Oct; 89(10): 2116-22. doi: 10.3382/ps.2010-00918. PMID: 20852102.

- Özkalp, B. (2012): Isolation and identification of Salmonellas from different samples. InTech Published, 1, 123-156.
- Park, S.Y.; Woodward, C.L.; Kubena, L.F.; Nisbet, D.J.; Birkhold, S.G. and Ricke, S.C. (2008): Environmental dissemination of foodborne Salmonella in preharvest poultry production: reservoirs, critical factors and research strategies. Crit. Rev. Environ.Sci. Technol., 2008. 38: 73-111.
- Shivaprasad, HL. and Barrow, P. (2008):
 Pullorum Disease and FowlTyphoid.
 In: Saif YM (ed.). Diseases of poultry,
 12th Edition. Blackwell Publishing,
 Ames, Iowa, pp. 620–636.

- Shivaprasad, HL. (2013): In: Swayne DE, ed. Diseases of Poultry. 13th ed. Ames, IA: John Wiley & Sons; 2013:706-713.
- Shivaprasad, H.L.; Cortes, P. and Crespo, R. (2006): Otitis interna (labyrinthitis) associated with Salmonella enterica arizonae in turkey poults. Avian diseases, 50(1), 135-138.
- Tracy, L.M.; Hicks, J.A.; Grogan, K.B.; Nicholds, J.A.; Morningstar-Shaw, B.R.; and Shariat, N.W. (2020): Molecular detection of Salmonella enterica subsp. arizonae by quantitative PCR. Avian Diseases, 64(3), 305-309.
- Wang, C. X.; Zhu, S.L.; Wang, X.Y.; Feng, Y.;
 Li, B.; Li, Y.G. and Liu, S.L. (2015):
 Complete genome sequence of Salmonella enterica subspecies arizonae str. RKS2983. Standards in Genomic Sciences, 10(1), 1-7.

الكشف عن نسبة الإصابة ببكتيريا السالمونيلا أريزونا في أنواع الدواجن المختلفة بمحافظة الوادي الجديد وحساسيتها وقدرتها المرضية في الديك الرومي

هشام محمد يحى محمد إحسان ، مصطفى عبد المطلب شحاته ، الشيماء رفيق سيد عثمان ، مروة محمد صفوت محمد توفيق ، مؤمن عبد العظيم محمد ، أحمد خلف عبد الحميد

E-mail: <u>hishamyahia@gmail.com</u> Assiut University website: <u>www.aun.edu.eg</u>

تعتبر عدوى الأريزنوزز في طيور الرومي الصغيرة واحدة من الأنماط المصلية للسالمونيلا التي تتميز بالإمراضية والنفوق العالي خلال تفشي المرض على نطاق واسع. تم جمع 250 عينة مشتبه بها من الطيور النافقة والمريضة من مختلف الأنواع في محافظة الوادي الجديد. تم إجراء العزل بالطرق القياسية من خلال وسائط الاستزراع المختبرية القياسية متبو عة باختبارات الكيمياء الحيوية. تم عمل اختبار حساسية لجميع المعزولات التي تم الحصول عليها باستخدام 6 مضادات بكتيرية لدراسة مساسية المعزولات للمضادات البكتيرية باستخدام طريقة أقراص المضادات الحيوية. تم عمل تجربه للكشف عن امراضية المعزولات من السالمونيلا أريزونا لصغار الديك الرومي البالغة من العمر 7 أيام تمت العدوى عن طريق الفم وداخل العين. ثلاثة وعشرون عزلة تم تحديدها على أنها سالمونيلا أريزونا بنسبة (2.2%) من طيور الديك الرومي والممان من مثلثة الأعمار. تم تسجيل حساسية عالية المعزولات الجنتاميسين يليه كوليستين سلفات ودوكسيسيكلين وتريميثوبريم / مختلف الأعمار. تم تسجيل معاسية عالية المعزولات للخبتاميسين يليه كوليستين سلفات ودوكسيسيكلين وتريميثوبريم / منطق مينوري مناز العدي معالية العزلات العزير ومي البالغة من العمر 7 أيام تمت العدوى عن طريق الفم وداخل العين. مختلف الأعمار. تم تسجيل حساسية عالية المعزولات للجنتاميسين يليه كوليستين سلفات ودوكسيسيكلين وتريميثوبريم / منطاميثوكسازول. تم تسجيل معاومة عالية المعزولات للفلور فينيكول والأموكسيسيلين. لوحظت اعراض وآفات ما بعد الوفاة في ملفاميثوكسازول. تم تسجيل مقاومة عالية للعزلات للفلور فينيكول والأموكسيسيلين. لوحظت اعراض وآفات ما بعد الوفاة في ملفاميثوكساز ال