Animal Health Research Institute, Port Said Laboratory for Food Hygiene.

A STUDY ON THE OCCURRENCE OF ESCHERICHIA COLI IN SOME BEEF PRODUCTS WITH SPECIAL REFERENCE TO ESCHERICHIA COLI 0157:H7.

(With 4 Tables)

By
ZIENAB I. SOLIMAN and AZZA A. EL-TABIY
(Received at 1/6/2006)

دراسة عن وجود ميكروب الإشيرشيا كولاى في بعض منتجات اللحوم مع الإشارة إلى عترة H7: 0157

زينب إبراهيم سليمان ، عزه على حسين التابعي

تعتبر منتجات اللحوم المصنعة من أهم المنتجات التي يقبل عليها المستهلك. وقد تتعرض لحوم الذبائح للتلوث بالميكروب القولوني أثناء الذبح وتختلط هذه البكتريا جيدا باللحم عند فرم وتجهيز اللحم لإعداد الأنواع المختلفة من منتجات اللحوم كالبرجر والسجق إلى غير ذلك من منتجات اللحوم المصنعة. ومع أن معظم عترات ميكروب الإشيرشيا كولاى غير ضارة إلا أن هناك بعض العنزات قد تشكل خطرا على صحة الإنسان. فعلى سبيل المثال العترة 7 O157:H تذكر دائما مرتبطة بحالات شديدة من التسمم الغذائي وخصوصا في منتجات اللحوم الغير مطهية جيدا وقد تؤدى العدوى إلى حدوث فشل كلوى وخاصة في الأطفال. لذلك تهدف الدراسة الحالية إلى عزل وتقييم انتشار الميكروب القولوني العترة H 7: 70157 المنابعة إلى جانب العترات الأخرى في بعض منتجات اللحوم المصنعة التي تباع في السوبر ماركت. تــم تجميع خمسة وعشرون عينة من كل من البرجر ، الفرانكفورتر، الكفتة، اللحم المفروم، والسجق. وقد أسفرت التحاليل عن وجود ميكروب الإشيرشيا كولاى بنسب ٥٦ % ، ٠٤% ، ٩٢٪ ، ٢٨٪ ، ٢٧٪ في عينات البرجر ، الفرانكفورتر، الكفتة، اللحم المفروم، والسجق على التوالي. كذلك تم عزل وتصنيف العترة O157 H:7 من ٦ (٤,٨ %) من اجمالي عدد العينات التي تم فحصها. كما تم تصنيف عترات أخرى من العينات الإيجابية وهــى 068 0119, 0113, 0119 وهــذا وقــد تم مناقشة خطورة وجود العسترة H 7: 1570 إلى جانب العترات الأخرى على الصحة العامة وكذلك أهم التوصيات بالنسبة لمستهاكي منتجات اللحوم المفرومة المصنعة والتي تتركز في الطهي الجيد وضمان وصول الحرارة إلى كافة الأجزاء الداخلية وليس السطح الخارجي وذلك للقضاء على الميكر وب.

SUMMARY

A total of one hundred and twenty five samples, twenty five of each beef burger, frankfurter, kofta, minced meat and sausage were collected from Port -Said markets. Samples were examined to isolate and evaluate the prevalence rate of *E. coli* O157:H7 and other *E. coli* serotypes. *E. coli* was detected in burger, frankfurter, kofta, minced meat and sausage samples at a rate of 56, 40, 92 68 and 72%, respectively. Six (4.8%) out of all 125 tested meat products samples were found to be contaminated with *E. coli* O157:H7, ten isolates of *E. coli* O157:H7 could be recovered. A total of 50 *E. coli* isolates recovered from positive samples were identified to serogroups, O55 (30%), O111 (22%), O113 (22%), O119 (16%), O68 (6%) and O126 (4%). The majority of *E. coli* serotypes recovered from the examined samples showed hemolytic activity. The public health significant of the isolated serogroups and consumer's safety were discussed.

Key words: Escherichia coli, beef products, E.coli 0157:H7.

INTRODUCTION

Many people enjoy beef burgers, sausages and other meat products, especially during the summer months. However, raw and improperly handled or cooked sausages and beef burgers can harbour harmful bacteria including *Escherichia coli*. The bacteria constituting the species *E. coli* are bacteria that normally live in the intestines of humans and animals. Although most strains are harmless, several are known to produce toxins that can cause diarrhea. The pathogenic groups includes enterotoxigenic *E. coli* (ETEC), enteropathogenic *E. coli* (EPEC), enterohemorrhagic *E. coli* (EHEC), enteroinvasive *E. coli* (EIEC), enteroaggregative *E. coli* (EAEC), diffusely adherent *E. coli* (DAEC). Of these, only the first 4 groups have been implicated in food or water borne illness(Levine, 1987 and Nataro and Kaper. 1998).

In recent years, it has become apparent that one can contract a rather serious bacterial gastro-enteritis by consuming undercooked ground beef. Scientists have identified a rare but dangerous type of *Escherichia coli*, *E. coli* O157:H7 that is responsible for this foodborne illness. Scientists believed that *E. coli* O157:H7 is a mutant strain that was created when a virus infected benign *E.coli* and gave it a string of DNA from *Shigella*, a bacterium that causes severe bloody diarrhea. In both *Shigella* and *E. coli* O157:H7, as few as 10 germs can cause illness;

by comparison, it takes about a billion Salmonella bacteria to make sick (Wong et al., 2000). E. coli O157:H7 was first recognized as a cause of illness in 1982 during an outbreak of severe bloody diarrhea; the outbreak was traced to under cooked burgers served from a fast food chains (Riley et al., 1983). Since 1983, an increasing in number of E. coli O157:H7 have been reported in association with consumption of improperly cooked ground beef (Cohen and Giannella, 1991 and Siegler et al., 1993).

The organism colonizes in the large intestine and produces one or more of the potent cytotoxins referred to as Shiga-like toxins (SLTs) (O'Brien and Kaper 1998). Although more than 60 *E. coli* serotypes produce SLTs, serotype O157:H7 is the predominant pathogen in the EHEC group and the one associated most frequently with human infections worldwide (Karmali, 1989). These toxins are responsible for severe hemorrhagic colitis in humans. In some persons, particularly children under 5 years of age and the elderly, the infection can also cause a complication called hemolytic uremic syndrome (HUS), in which the red blood cells are destroyed and the kidneys fail, about 2%-7% of infections lead to this complication (Doyle, 1991).

In a view of the importance of *E. coli* O157:H7 from a food safety stand point, this study was planned to investigate the presence of this agent and other pathogenic *E. coli* serotypes among some selected meat products. The public health significant and consumer's safety were discussed.

MATERIALS and METHODS

One hundred and twenty five samples, twenty-five of each beef burger, frankfurter, minced meat, kofta and sausage were collected from Port- Said markets.

The frozen samples were thawed in their original containers in a refrigerator at 2-5°C. Twenty-five grams of each sample were homogenized with 225 ml of tryptone phosphate broth as a preenrichment fluid then incubated for 4-6 hours at 37°C. (Mehlman and Lovett, 1984). Two Mossel's enteric enrichment broth tubes (10 ml) were inoculated each by 1 ml from the pre-enrichment medium. One tube was incubated at 44°C for 24 hours to permit the growth of pathogenic *E.coli*, other than serovar O157. The other tube was incubated at 37°C for 24 hours to permit the growth of *E. coli* O157:H7 as well as other serovars unable to grow at high (44°C) temperature (Mehlman and Romero, 1982). Dilutions of culture in tryptone phosphate broth with

peptone water (0.1%) to 10⁻⁶ were prepared. About 0.1 ml obtained from appropriate dilution were inoculated in MacConkey Sorbitol agar (MACS) and Eosin methylene blue agar (EMB) as double parallel by using spread -plating. The plates were incubated at 37°C for 24 hours. Randomly selected white and colorless sorbitol negative colonies were picked from MACS and streaked separately onto MACS supplemented with cefixime- tellurite (CT, Difco) (CT- MACS) and onto EMB to purify the colonies. The plates were incubated at 37°C for 24 hours (FDA, 2002). Morphological and biochemical tests were applied to colorless or neutral /gray with smoky center and 1-2 mm diameter sorbitol negative colonies on CT- MACS and to metallic green colored, smooth sided colonies on EMB according to Quinn et al. (2002). The isolates were identified serologically by the slide agglutination test using diagnostic polyvalent and monovalent E.coli O antisera and H 7 antisera (Escherichia coli antisera, Denka Seiken Co., Ltd., Tokyo, Japan) following the manufacturer's specification.

Hemolysin production (Beutin et al., 1989)

E. coli isolates were inoculated onto blood agar plates containing sheep blood (5%) and incubated at 37°C for 24 hours. The plates were examined for the presence of haemolysis.

RESULTS

Table 1: Prevalence rate of *Escherichia coli* in the examined meat products samples (n=25 of each).

Meat products	Positive samples	% of Positive samples 56 40 92		
Beef burger	14			
Frankfurter	10			
Kofta	23			
Minced meat	17	68		
Sausage	18	72		

Table 2: Prevalence of *E. coli O157*: H7 among the examined meat products (n=25 of each).

Meat products	Positive samples for serovar O157:H7			
Tital products	No.	%		
Beef burger	2	8		
Frankfurter	0	0		
Kofta	1	4		
Minced meat	3	12		
Sausage	0	0		
Total	6	4.8		

Table 3: Serovars of *E. coli* isolates recovered from the examined meat products samples.

Serovar	055		011	I	0113	3	0119	7	O68		012	16	
source	No	%	No	%	No	%	No	%	No	%	No	%	
Beef burger	4	8	3	6	4	8	3	6	1	2	2	4	
Frankfurter	1	2	2	4	1	2	0	0	1	2	0	0	
Kofta	5	10	4	8	3	6	3	6	0	0	0	0	
Minced meat	3	6	2	4	1	2	1	2	1	2	0	0	
Sausage	2	4	0	0	2	4	1	2	0	0	0	0	
Total	15	30	11	22	11	22	8	16	3	6	2	4	

NB: Percentage was calculated according to the total number of the isolates (50)

Table 4: Hemolytic activity of *E. coli* isolates recovered from the examined meat products samples.

E. coli serovars	No. of isolates	Hemolytic activity		
	No. of isolates	No.	%	
O157: H7	10	10	100	
O55	15	15	100 81.8	
O111	11	9		
O113	11	7	63.6	
O119	8	8	100	
O68	3	0	0	
O126	2	0	0	
Total	60	49	81.7	

DISCUSSION

Most enteropathogenic *E. coli* outbreaks have been blamed on ground beef and other meat products such as beef burger, and hot dog (Desmarchelier and Grau, 1997). The present investigation was carried out to evaluate the prevalence of *E. coli* O157: H7 and other *E. coli* serotypes among selected types of meat products.

The overall incidence of *E. coli* in different samples was recorded in Table (1), *E. coli* were recovered from burger, Frankfurter, kofta, minced meat and sausage samples at a rate of 56, 40, 92, 68 and 72%, respectively. In this concern, prevalence of *E. coli* from meat and meat products ranging from 30% to 76% have been reported by Doyle and Schoeni, (1987), Gallas *et al.*, (2002) and Gad El-Said *et al.*, (2005). This contamination rate of the present samples indicates unhygienic

practices prevailed in slaughter. Cattle are a major reservoir of these groups of bacteria and ground beef have been the major vehicle of *E. coli* transmission. During slaughter process, meat may become contaminated by fecal contamination during evisceration and through skin or hide during dressing (Desmarchelier, 1997 and Rice *et al.*, 1997). When the meat is ground, fecal organisms on the outside of the meat are mixed throughout the ground beef. Also contamination of meat probably occurs during processing. In this respect, Read *et al.* (1990) reported that ground beef meat- processing plants were heavily contaminated with verocytotoxin *E. coli*. In addition, *E. coli* is an indicator of food safety for dehydrated, frozen and refrigerated food, as *E. coli* does not survive well under such condition (Mossel *et al.*,1979). Therefore, its presence might indicate poor temperature control.

Escherichia coli O157: H7, predominantly originated from beef, is a significant pathogen to the public health and thus, need to be vigorously surveyed in meat products. Lake of sorbitol fermentation within 24 hours has been considered a stable phenotypic character of E. coli O157: H7 therefore; MACS was used for differentiation of E. coli O157: H7 from other enteric bacteria (March and Ratnam, 1986).

Results of biochemical and serological identification of sorbitol negative E. coli isolates revealed that six (4.8%) out of all 125 meat products examined were found to be contaminated with E. coli O157:H7 (Table 2), three (8%) out of 25 minced meat samples, two (8%) out of 25 burger samples and one (4%) out of 25 kofta samples. The exact contamination rate may be higher than stated here due to the low isolation rate of the culture methods compared to other immunological and genetical methods. Considerably higher isolation rates of E. coli O157:H7 than in this study have been reported elsewhere. In South Africa, it was isolated from a total of 74.5% and in Malaysia from 36% of beef samples (Vorster et al., 1994 and Radu et al., 1998). On the other hand, in some studies beef and beef samples have found to be free (Junghannss et al., 1996, Simmons, 1997 and Uhitil et al., 2001). In another studies it was isolated at low contaminated rate, Pai et al., (1984) reported the presence of E. coli O157:H7 in 5 out of 17 beef samples. In USA, E. coli O157:H7 was isolated from six (3.7%) out of 164 beef samples (Doyle and Schoeni, 1987), in India, Dutta and Deb, (2000) isolated E. coli O157:H7 from two (9%) out of 22 minced beef samples. Also in turkey, Baran and Gulmez, (2005) isolated E. coli O157:H7 from three (6%) of ground beef samples. Positive isolation of E. coli O157:H7 from beef samples in Egypt was reported by Tanios et

al., (2002) from two (6.7%) of minced meat samples and by Gad El-Said et al., (2005) with a rate of 3.95% from meat samples.

Enterohemorrahagic Escherichia coli O157: H7 is an important foodborne pathogen, its presence even at low rate (4.8%) in the present study may constitutes dangerous beef products. The ability of E. coli O157:H7 to withstand the acidic conditions encountered in various foods have generally suggested that passage through the stomach would be insufficient to inactivate the pathogen. (Naim et al., 2003). In addition, the organism always enters the digestive system within a food matrix, Waterman and Small (1998) postulated that high protein content in food (such as ground beef and boiled egg white) might protect enteric bacteria against the killing effect of gastric acids. The data from epidemiological investigations indicated that as few as 10 to 100 cells of E. coli O157:H7 per g of raw ground beef were sufficient to cause illness (Abdul-Raouf et al., 1993). Moreover, Wong et al. (2000) believed that treatment with antibiotics is contraindicated for E. coli O157 poisoning, since it is when the bacteria die, they release the toxins which produce hemolytic uremic syndrome (HUS), for which there is no cure.

Symptoms of *E. coli* O157 infection include bloody and nonbloody diarrhea, vomiting, and abdominal cramps. Illness resolves typically within 7-10 days .A subset of patients, particularly the young and the elderly, will develop HUS, characterized by microangiopathic hemolytic anemia, thrombocytopenia, and renal failure (Russell *et al.*, 2000). In the United States, hemolytic uremic syndrome is the principal cause of acute kidney failure in children, and *E. coli* O157: H7 causes most cases of hemolytic uremic syndrome (Besser *et al.*, 1999). An estimated 73,480 people a year are infected with *E. coli* O157:H7 and about 600 of those cases are fatal, according to the federal Centers for Disease Control and Prevention (Wong *et al.*, 2000). These illnesses and deaths were factors that began changing policy towards foodborne disease. The Food Safety and Inspection Service, declared that raw ground beef contaminated with *E. coli* O157 is adulterated and must be further processed to kill the pathogen or be destroyed (FDA., 2000).

While *E. coli* O157: H7 is the most renowned Shiga toxin-producing *E. coli* (STEC), over 200 different types of STEC have been documented in meat and animals, at least 60 of which have been linked with human disease. A number of studies have suggested that non-O157 STEC are associated with clinical disease, and non-O157 STEC are present in the food supply (Acheson, 2000).

Regarding to other serogroups, as shown in Table (3), O55, O111, O113 and O119 were the most prevalent serotypes recovered from the examined samples with an incidence of 30, 22, 22, and 16% respectively, followed by O68 (6%), O126 (4%). Most of the isolated serotypes are usually associated with many cases of foodborne outbreaks and multiple sporadic cases in different part of the world. In this concern, Anathan and Subramaniam, (1995) isolated E. coli belonging to serotypes O111 from cases of persistent diarrhea in young children. Enteropathogenic E. coli belonging to serotypes O111, O103 and O55 were isolated from patients suffering from bloody diarrhea, which may be accompanied by HUS (Desmarchelier, 1997). Non-O157 STEC, such as O111 has caused large outbreaks and HUS in the United States and other countries. (Acheson, 2000). Moreover, Hussein and Omaye, (2003) found that the serogroups belonging to O26, O113, O111, O119 and O166 have caused approximately 30% of the hemolytic uremic syndrome (HUS) in US.

Blood haemolysis is one of character of virulent *E. coli* (Stephen et al., 1985).

Ten isolates identified serologically as *E. coli* O157: H7 were tested for hemolysis production using sheep blood. All tested isolates were haemolytic. Moreover, the majority of *E. coli* isolates other than O157: H7 isolated from the examined samples showed haemolytic activity (Table 4). In this respect, Adesiyun *et al.*, (1997) reported that from 94 *E. coli* isolates tested for haemolysis 13.8 % were haemolytic. Meanwhile, Gad El-Said *et al.*, (2005) stated that 81.58 % of *E. coli* isolates recovered from meat samples showed haemolytic activity.

The productions of haemolysin have a potential role in virulence of hemolytic *E. coli*. Therefore, contamination of meat products with *E. coli* O157:H7 and other *E. coli* serotypes may results in problems for consumers. There is a close association between enterohaemolysin production and SLT production (Beutin *et al.*, 1998). Moreover, the genes involved in enterohaemolysin production were carried on the EHEC plasmid (Scotland *et al.*, 1990).

The risk of contamination of raw meat products with *E. coli* O157:H7 and other pathogens constitute a major problem for human. The low infective dose *E. coli* O157:H7 present a major threat. Hemolytic uremic syndrome, a disease caused mostly by *E. coli* O157:H7 may cause sever kidney diseases and/or failure among children. The main means of combating this organism are good food hygiene covering activities on farm, in abattoir and minced beef

industries. However, until E. coli can be eliminated from meat processing systems, consumers should protect themselves by using safe food practices and advice for those who eat ground beef. Frozen ground beef should be thawed in the refrigerator rather than at room temperature. While thawing and preparing ground beef, raw meat must be separated from ready-to-eat foods. It is not enough to merely brown the outside of a burger; and other meat products .Ground beef should be cooked thoroughly to an internal temperature of at least 160° F (71° C). food safety experts recommends that consumers use a meat thermometer to cook ground beef to ensure that internal temperatures are high enough to kill bacteria. To reduce the risk for cross-contamination, consumers should use soap and hot water to wash hands, utensils, and other surfaces that might have been exposed to raw or undercooked ground beef and other meat products. In addition, consumers should be aware from under cooked burgers and other meat products served from fast food restaurants

REFERENCES

- Abdul-Raouf, U.M.; Beuchat, L.R. and Ammar, M.S. (1993): Survival and growth of Escherichia coli O157:H7 in ground beef as affected by pH, acidulant, and temperature. Appl. Environ. Microbiol., 59: 2364-2368.
- Acheson, D.W.K. (2000): How does Escherichia coli O157: H7 testing in meat compare with that we are seeing clinically. J. Food Protect., 63 (6): 819 821.
- Anathan, S. and Subramaniam, M.A. (1995): Microbial etiology and assessment of immunological status of children persistent diarrhea. J. Comm. Dis., 27(3): 193-195.
- Adesiyun, A.A.; Webb, L.A.; Romaine, H. and Kaminjolo, J.S. (1997):

 Prevalence and characteristics of strains of Escherichia coli
 isolated from milk and feces of cows on dairy farms in
 Trinidad J. Food Protect., 60 (10): 1174-1181.
- Baran, F. and Gulmez, M. (2005): Escherichia coli O157:H7 in the ground beef and chicken drumsticks. Internet Journal of Food Safety., 2: 13-15.
- Besser, R.E.; Griffin, P.M. and Slutsker, L. (1999): Escherichia coli O157:H7 gastroenteritis and the hemolytic uremic syndrome: an emerging infectious disease. Annu. Rev. Med., 50: 355-367.

- Beutin, L.; Montenegro, M.A.; Orskov, I.; Orskov, F.; Prada, J; Zimmermann, S. and Stephen, R. (1989): Close association of verotoxin (Shiga like toxin) production with enterohaemolysin production in strains of E. coli. J. Clin. Microbiol., 27 (11): 2559-2564.
- Beutin, L.; Zimmermann, S. and Gleier, K. (1998): Human infections with shiga toxin-producing Escherichia coli other than O157 in Germany. Emerg. Infect. Dis., 4: 635-639.
- Cohen, M.B. and Giannella, R.A. (1991): Hemorrhagic colitis associated with Escherichia O157:H7. Adv. Intern. Med., 37:173-195.
- Desmarchelier, R.M. (1997): Enterohemorrhagic E. coli. The Australian Perspective J. Food Protec., 60 (11): 1447-1450.
- Desmarchelier, R.M. and Grau, F.H. (1997): Escherichia coli
 .Foodborne Microorganisms of Public Health Significance. A.
 D. Hocking, G. Arnold, I. Jenson, K. Newton and P. Shuterland
 (ed.) P.231-264. Australian Institute of Food Science and
 Technology Inc., Sydney, Australia.
- Doyle, M.P. (1991): Escherichia coli O157:H7 and its significance in foods. Int. J. Food Microbiol., 12:289–302.
- Doyle, M.B. and Schoeni, J.L. (1987): Isolation of Escherichia coli O157:H7 from retail meats and poultry. Appl. Environ. Microbiol., 53(10): 2394-2396.
- Dutta, S. and Deb, U.K. (2000): Isolation of Escherichia coli O157:H7 strains from dairy cattle and beef samples marketed in Calcutta, India. J. Med. Micobiol., 49: 765-767.
- FDA "Food and Drug Administration" (2000): U. S. Department of Agriculture, U. S. Environmental Protection Agency. National food safety initiative.
- FDA "Food and Drug Administration" (2002): U.S. Food and Drug Administration Center for Food Safety & Applied Nutrition, Bacteriological Analytical Manual Online, Chapter 4A, Diarrheagenic Escherichia coli.
- Gad El- Said, W.A.; El- Jakee, J.K.; Kandel, M.M. and El- Shabrawy, M.A. (2005): Presence of Escherichia coli O157:H7 in raw milk and meat samples. J. Egypt. Vet. Med. Assoc., 65(3): 341-350.
- Gallas, N.; Ben Aissa, R.R.; Attia, A.T.; Bahri, O. and Boudabous, A. (2002): Isolation and characterization of shiga toxin- producing Escherichia coli from meat and dairy products. Food Micobiol.,19: 389-398.

- Hussein, H.S. and Omaye, S.T. (2003): Introduction of food safety concerns of verotoxin producing Escherichia coli. Exp. Biol. Med. (Maywood)., 228 (4): 331-332.
- Junghannss, U.; Winterfeld, S. and Freerksen, R. (1996): Incidence of Enterohaemorrhagic colitis E. coli in meat. Gesundheitswesen., 58: 482-484.
- Karmali, M.A. (1989): Infection by Verocytotoxin-producing Escherichia coli. Clin. Microbiol. Rev., 2: 15-38.
- Levine, M.M. (1987): Escherichia coli that cause diarrhea enterotoxigenic, enteroinvasive, enterohemorrhagic, and enteroadherent. J. Infect. Dis., 155:377-389.
- March, S.B. and Ratnam, S. (1986): Sorbitol MacConkey medium for detection of Escherichia O157:H7 associated with Hemorrhagic colitis. J. Clin. Microbiol., 23: 869-872.
- Mehlman, I.I. and Lovett, J. (1984): Escherichia coli in FDA Bacteriological Analytical Manual, 6th ed. Assoc. of Official Analytical Chemists: Arlington.
- Mehlman, I.I. and Romero, A. (1982): Escherichia coli recovery from foods. Food Technol., 36 (3): 73-79.
- Mossel, D.A.A.; Eelderink, L.; Koopmans, M. and Van Rossens, F. (1979): Influence of carbon source, bile salts and incubation temperature on recovery of Enterobacteriaceae from food using MacConkey type agar., J. Food Protect 24: 470-475.
- Naim, F.; Messier, S.; Saucier, L. and Piette, G. (2003): A model study of Escherichia coli O157: H7 survival in fermented dry sausages—influence of inoculums preparation, inoculation procedure and selected process parameters. J. Food Prottect 66: 2267-2275.
- Nataro, J.P. and Kaper, J.B. (1998): Diarrheagenic Escherichia coli. Clin. Microbiol. Rev., 11:132-201.
- O'Brien, A.D. and Kaper, J.B. (1998): Shiga toxin-producing Escherichia coli: yesterday, today, and tomorrow, p. 1-11. In J. B. Kaper and A. D. O'Brien (ed.), Escherichia coli O157:H7 and other Shiga toxin-producing E. coli strains. American Society for Microbiology, Washington, D.C.
- Pai, C.H.; Gordon, R.; Sims, H.V. and Bryan, L.E. (1984): Sporadic cases of Hemorrhagic colitis associated with Escherichia coli O157:H7. Ann. Intern. Med., 101:738-742.

- Quinn, P.J.; Markey, B.K.; Carter, M.E.; Donelly, W. J. C. and Leonard, F.C. (2002): Veterinary Microbiology and Microbial Diseases. 1st Iowa State University Press Blackwell Science.
- Radu, S.; Rusul, G.; Morigaki, N.; Asai, Y.; Kim, J.; and Nishibuchi, M. (1998): Detection of Escherichia coli O157:H7 in the beef marketed in Malaysia. Appl. Environ. Microbial., 64 (3): 1153-1156.
- Read, S.C.; Gyles, C.L.; Clark, R.C.; Liar, H. and McEwen, S. (1990):

 Prevalence of verocytotoxigenic Escherichia coli in ground beef, pork and chicken in Southwestern Ontario. Epidemiol. Infec., 105: 11-20.
- Rice, D.H.; Ebel, E.D.; Hancock, D.D.; Besser, T.E.; Herriott, D. and Carpenter, L.V. (1997): Escherichia coli O157:H7 in cull dairy cows on farm and at slaughter .J. Food Protect 60 (11): 1386-1387.
- Riley, L.W.; Remis, R.S.; Helgerson, S.D.; McGee, H.B.; Wells, J.G.; Davis, B.R.; Herbert, R.J.; Olcott, E.S.; Johnson, L.M.; Hargett, N.T.; Blake, P.A. and Cohen, M.L. (1983): Hemorrhagic colitis associated with a rare Escherichia coli serotype. N. Engl. J. Med., 308:681-85.
- Russell, J.B.; Diez-Gonzalez, F. and Jarvist, G.N. (2000): Effects of diet shifts on Escherichia coli in cattle. J. Dairy Sci., 83 (4): 863-872.
- Scotland, S.M.; Willshaw, G.A.; Smith, H.R. and Rowe, B. (1990):
 Properties of strains of Escherichia coli O26: H11 in relation to their enteropathogenic or enterohemorrhagic classification. J. Infect. Dis., 162 (5): 1069-1074.
- Siegler, R.L.; Griffin, P.M. and Barrett, T.J. (1993): Recurrent hemolytic uremic syndrome secondary to Escherichia coli O157:H7 infection. Pediatrics; 91: 666-68.
- Simmons, N.A. (1997): Global perspectives on Escherichia coli O157:H7 and other verocytotoxic E. coli spp. J. Food Protect 60 (11): 1463-1465.
- Stephen, J.; Cavalieri, G.; and Synder, I. (1985): Escherichia coli α Haemolysin: characteristics and probable role in pathogenicity. Microbiological Reviews, Dec. 48 (4): 326-343.
- Tanios, A.I.; Shaaban, A.I.; Abd –El-aty, I. and El-Shernoby, R. (2002): Serological features of pathogenic E. coli in meat and meat products. J. Egypt. Vet. Med. Assoc., 62 (3): 245-253.

- Uhitil, S.; Jaksic, T.; Petrak, T. and Botka-Petrak, K. (2001): Presence of Escherichia coli O157:H7 in ground beef and baby beef meat. J. Food Protect 64 (6): 862-864.
- Vorster, S.M.; Greebe, R.P. and Nortje, G.L. (1994): Incidence of Staphylococcus aureus and Escherichia coli in ground beef, broilers and processed meat in Pretoria, South Africa. J. Food Protect 57: 305-310.
- Waterman, S.R. and Small P.L.C. (1998): Acid-sensitive enteric pathogens are protected from killing under extreme acidic conditions of pH 2.5 when they are inoculated onto certain solid food sources. Appl. Environ. Microbiol., 64:3882-3886.
- Wong, S.C.; Jelacic, S.; Habeeb, R.L.; Watkins, S.L. and Tarr, p.I. (2000): The risk of the hemolytic uremic syndrome after antibiotic treatment of Escherichia coli O157:H7 infections. N. Engl. J. Med., 5: 23-28.