

IMPROVEMENT OF THE FERILITY AND SOME BIOCHEMICAL EFFECTS OF AFLATOXIN ON MALE RABBIT BY GENSING

(With 4 Tables and 4 Figures)

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**الدور الواقى للجنسج في تقليل التأثير الضار للأفلاتوكسين على الخصوبة
وبعض الوظائف البيوكيميائية في ذكور الأرانب**

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أجرى هذا البحث لدراسة تأثير الأفلاتوكسين الذي يكون موجودا في العلائق على الخصوبة وبعض القياسات البيوكيميائية لذكور الأرانب البلدي وبيان الدور الوقائي للجنسج لمنع تلك التأثيرات. في هذه الدراسة تم استخدام عدد ٦٠ من ذكور الأرانب البلدي في مزرعة خاصة بمحافظة الشرقية تتراوح أعمارها ٨ - ١٠ شهور ووزنها يتراوح بين ٢,٥ - ٣ كجم وقسمت هذه الأرانب إلى أربع مجموعات متساوية (١٥ أرنب في كل مجموعة) الأولى تركت بدون اي علاجات كمجموعة ضابطة والثانية تم إضافة الأفلاتوكسين في العليقة بمقدار ١ ملجم /كجم عليقة يوميا لمدة شهرين الثالثة تم إعطاء مستخلص الجنسج للأرانب بعد إذابة في مياه مقطرة عن طريق أنبوبة اللي المعدي بمقدار ٢٠٠ ملجرام /كجم عليقة يوميا لمدة شهرين والرابعة تم إضافة الأفلاتوكسين والجنسج يوميا لمدة شهرين. تم أخذ عينة دم وسائل منوي من كل حيوان عند اليوم الأول، ٣٠ و ٦٠ من نهاية استخدام الأفلاتوكسين والجنسج وذلك لفصل المصل وذلك لقياس هرمون التستستيرون وبعض المؤشرات البيوكيميائية. وأظهرت النتائج بعد تحليلها إحصائيا أن الأفلاتوكسين أدى إلى نقص معنوي في تركيز الحيوانات المنوية ومعدل الحركة وعدد الحيوانات المنوية الحية كما أدى إلى زيادة نسبة العيوب الشكلية في الحيوانات المنوية الحية. أما الجنسج بالجرعة التي تم استخدامها أحدث زيادة معنوية في تركيز الحيوانات المنوية ومعدل الحركة وعدد الحيوانات المنوية الحية كما أدى إلى نقص معنوي في نسبة العيوب الشكلية الكلية في الحيوانات المنوية الحية. كما لوحظ إن الأفلاتوكسين والجنسج معا أديا إلى زيادة غير معنوية في تركيز الحيوانات المنوية ومعدل الحركة وعدد الحيوانات المنوية الحية ولكن نسبة العيوب الشكلية الكلية في الحيوانات المنوية الحية لم تتأثر. الأفلاتوكسين أدى إلى وجود نقص معنوي في معدل هرمون التستستيرون والبروتين الكلى وزيادة معنوية في معدل كلا من الترانس امينيزز (ALT-AST), والفوسفاتيز القاعدي في مصل الدم. أما الجنسج بالجرعة التي تم استخدامها أحدث زيادة معنوية في هرمون التستستيرون والبروتين الكلى , كما وجد أن الترانس امينيزز (-ALT AST), والفوسفاتيز القاعدي لم تتأثر. كما لوحظ إن تلك المؤشرات البيوكيميائية والتستستيرون لم تتأثر باستخدام الأفلاتوكسين والجنسج معا. نستخلص من هذه الدراسة أن

الأفلاتوكسين أحدث تأثيرات عكسية على الخصوبة الذكري، هرمون التستوستيرون، بعض القياسات البيوكيميائية في ذكور الأرانب ولكن استخدام الجينسج أدى إلى تلافى تلك التأثيرات. لذلك ينصح باستخدام الجينسج في مزارع الأرانب لكي يعادل تأثير الأفلاتوكسين في العلائق إن وجد وكمنشط للخصوبة والوظائف الحيوية للأرانب في المزرعة.

SUMMARY

The present study was carried out to investigate the effect of aflatoxin contaminated feed on male fertility and some biochemical parameters in male rabbits and modulating this effect by using gensing. A total 60 of apparently healthy adult male baladi rabbits (8-10 month of age) and 2.5-3 kg b.wt., were divided into four equal groups, 1st group was left without treatment as control, 2nd group was fed on a ration containing 1 mg aflatoxin /kg for 60 successive days, 3rd group received 200 mg/kg body weight of gensing extract dissolved in saline solution orally through mouth tube for 60 successive days and 4th group was fed a ration containing 1 mg aflatoxin /kg and received 200 mg/kg by using mouth tube for 60 successive days. Five rabbits from each groups were slaughtered at 1st, 30th and 60th days post medication. Blood and semen samples were collected from control and treated rabbits. Blood samples were used for biochemical analysis. Semen samples were collected from tail of the epididymis to evaluate semen character. Aflatoxin elucidated significant reduction on sperm cells concentration, progressive motility and live sperm but increased total abnormality percent at 1st and 30th days post aflatoxin medication. Gensing induced significant increase in sperm cells concentration, progressive motility, live sperm and decreased total abnormality percent at 1st and 30th days post treatment. Rabbits received both aflatoxin and gensing induced non significant effect on sperm cells concentration, progressive motility, live sperm and total abnormality percent. Aflatoxin induced significant decrease in testosterone hormone level, weight of secondary sex organs (testis, seminal vesicle and prostate gland), total protein and significant elevation in serum transaminases (AST-ALT) and alkaline phosphatas at 1st and 30th days post feeding on aflatoxin. Gensing induced significant increase in the level of serum total protein, testosterone hormone but ALT, AST and alkaline phosphatas non significantly effect. Aflatoxin and gensing induced non significant effect in the above biochemical parameters. It could be concluded that aflatoxin induced many reversible alteration in male fertility, hormonal and some biochemical parameters as they returned to their normal values 60th day after stopping aflatoxin

addition. Gensing minimize or reduce the alteration in fertility and biochemical changes induced by aflatoxin.

Key words: Rabbit, feretility, aflatoxin

INTRODUCTION

Aflatoxins are toxic metabolites produced by fungi of *Aspergillus* species. They are produced by many strains of *Aspergillus flavus*, *Aspergillus parasiticus* and *Penicillin puberulum* (Peterson, *et al.*, 2001). There are four generally aflatoxin B1, B2, G1 and G2 Aflatoxin B1 is the more toxic. (Ciegler and Bennet, 1980). The susceptibilily of animals to aflatoxins varies considerably, depending on sex, age, species and nutrition (Eaton and Groopman, 1994). Aflatoxin also induce anemia,poor digestion,inhibition of protein synthesis,lipid peroxidation and oxidative DNA damage (Verma and Nair, 2001). Aflatoxicosis reduce the male fertility (Chao, 1991).

Many studies were attempted to reduce aflatoxin in the contaminated food by using numerous physical, chemical and/or biological technique (Samarajeewa *et al.*, 1991). Some herbal food supplements were found to reduce the toxic effect of aflatoxin through its antioxidant effect (El-Seidy *et al.*, 2002). Also some medicinal herbs has a protective action against free radical (Ahmed *et al.*, 2000).

Panax gensing is one of the most valued medicinal herbaceous plant belonged to family Araliaceae (Kamel and Hoda, 2006). It is herbal root, has a wide pharmacological action in the clinical practice (Chong and Oberholzer, 1989). This plant contain many valuable ingredients such as saponins (known as panaxosides or gensingosid, vitamin A, B6, mineral as zinc, antioxidant, peptides, fatty acids, polysaccharide, alcohol and cholesteryl ester transfer protein inhibitors (Huang, 1999). The various forms of gensing appear to be non toxic (Hess *et al.*, 1982). Gensing decreases nitric acid content and nitric oxide synthase activity (play a role in accelerating senility) in the cerebral cortex in rats (Li *et al.*, 1997). Gensing improves the survival rate and sperm quality in guinea pigs (Kim *et al.*, 2004).

The objective of the present work was to evaluate the effect of dietary suppl-ementation of aflatoxin on the male fertility and some biochemical parameters in rabbit and modulating this effect by using gensing.

MATERIALS and METHODS

1- Drug:-

Gensing Extract (Korean red Gensing extract) was obtained from Pharco Pharmaceuticals, Alexandria, Egypt in capsules and each capsule contain 100 mg of gensing extract.

2-Animals:-

The present investigation was carried out on sixty male rabbits 8 months old and about 2.5-3 kg body weight. Rabbits were obtained from private rabbit farm in Sharkia Province. Rabbits were housed under hygienic condition, feed comm-ercial pellets (Table 1) and watered adlibitum during the experimental period.

Table 1: Ingredients of experimental diet.

Component	%
Barly	25
Bran	25
Soya bean meal 44 % protein	3
Sun flower meal 24 % protein	24.55
Berseem hay	15
Soya oil	3
Bone meal	2
Lime stone	1
Sodium chloride	1
Methionine	0.15
Vitamins/ minerals premix	0.3

3-Experimental design:

Rabbits were divided into four equal groups (15 rabbits each). First group left without treatment as control group, 2nd group was fed on a ration containing 1 mg aflatoxins / kg ration for 60 successive days, 3rd group received 200 mg of gensing extract /kg body weight extract, that dissolved in saline solution then by using mouth tube introduce the extract daily for 60 days (Choi *et al.*, 1999), and 4th group was fed on a ration containing 1 mg aflatoxins /kg ration and 200 mg of gensing extract / kg b.wt.for 60 successive days. Five rabbits from each groups were slaughtered at 1st, 30th and 60th days post medication. Blood samples from control and treated rabbits were taken for obtained clear serum for biochemical analysis and Semen sample were taken from tail of the epididymis to evaluate semen characters.

A) Examination of epididymal sperm

After rabbits have been slaughtered the epididymal content of each rabbit was collected and squeezed gently in sterile watch glass containing 1ml sodium citrate solution 2.9% to estimate the percentage of progressive motility, sperm cell concentration / mm³, alive sperm

percent and percentage of total sperm abnormalities according to the method described by Bearden and Flaquary (1980).

B) Blood samples hormonal and biochemical assays

One blood samples from control and treated rabbits were taken at 1st, 30th and 60th days post medication and collected in test tubes during animal slaughtering and left to clot at room temperature then centrifugated for about ten minutes at 3000 r.p.m to obtain clear serum for determination of testosterone hormone by radioimmunoassay according to the method described by Wilson and Foster (1992), aspartate aminotransferase (AST), alanine aminotransferase (ALT) activities according to the methods described by Reitman and Frankel (1957), serum alkaline phosphatase according to Kind and King(1954) and total protein according to Doumas (1975).

C) Sex organs weight

Following slaughtering of the male rabbits testis and secondary sex organs (seminal vesicle and prostate gland) were dissected out and weighted at 1st, 30th and 60th days post medication.

4-Statistical analysis:-

The obtained results were statistically by Student t test were explained by Petrie and Watson (1999)

RESULTS

The obtained results were tabulated in Tables 2, 3, 4 and Fig A, B, C and D

Table 2: Effect of aflatoxin (1 mg /kg BW) gensing(200 mg /kg BW) either alone or together on sperm cell concentration (x106 spz/ml), sperm motility(%), live sperm (%) total sperm abnormality (%) and Testosterone hormone (ng/ml) at 1st, 30th and 60th days post medication for two months in male baladi rabbits (n=5)

parameter	Healthy rabbit (control)	1 st days			30 th days			60 th days		
		AF	GE	AF+GE	AF	GE	AF+GE	AF	GE	AF+GE
Spreem cell concentration	2.95±0.28	1.83±0.16**	3.92±0.19*	3.16±0.23	2.06±0.12*	3.71±0.04*	3.04±0.16	2.53±0.18	3.12±0.23	2.98±0.17
Sperm motility	82.73±1.83	70.31±2.40**	90.52±1.76*	76.90±1.73	75.61±1.93*	88.70±1.72*	78.52±1.96	79.63±1.65	83.71±1.98	80.25±1.47
Live Sperm	85.23±1.53	70.12±3.20**	91.32±1.36*	83.0±1.46	78.16±1.99*	89.15±0.86*	84.20±1.31	82.92±1.59	85.13±1.26	85.10±1.43
Total Abnormality	12.31 ± 0.32	17.42±1.37**	9.16±0.22*	13.27±0.68	15.03±0.42	10.36±0.28	12.59±0.68	13.05 ± 0.82	11.63±0.73	12.05±0.93
Testosterone	3.13±0.53	1.20±0.14**	5.26±0.16**	3.60±0.24	2.02±0.25*	4.49±0.13*	3.3±0.32	2.92±0.37	3.06±0.42	4.32±0.26

* Significant at P < 0.05

** Significant at P < 0.01

Table 3: Effect of aflatoxin (1 mg /kg BW) gensing(200 mg /kg BW)

either alone or together on weight of testis (gm), seminal vesicle (gm) and prostate gland (gm) at 1st, 30th and 60th days post medication for two months in male baladi rabbits (n=5)

Parameter	Healthy rabbit	1 st days			30 th days			60 th days		
		AF	GE	AF+GE	AF	GE	AF+GE	AF	GE	AF+GE
Testis	2.37± 0.21	2.02± 0.19	2.09± 0.21	2.23± 0.25	2.61± 0.19	2.69± 0.14	2.9± 0.19	2.31± 0.25	2.39± 0.19	2.35± 0.17
Seminal vesicle	1.92± 0.21	1.78± 0.24	1.83± 0.19	1.89± 0.22	2.08± 0.41	2.12± 0.38	2.1± 0.21	1.85± 0.25	1.88± 0.22	1.90± 0.18
Prostate	0.56± 0.07	0.47± 0.18	0.53± 0.12	0.58± 0.09	0.59± 0.11	0.57± 0.09	0.5± 0.03	0.51± 0.05	0.53± 0.06	0.52± 0.08

*Significant at P < 0.05

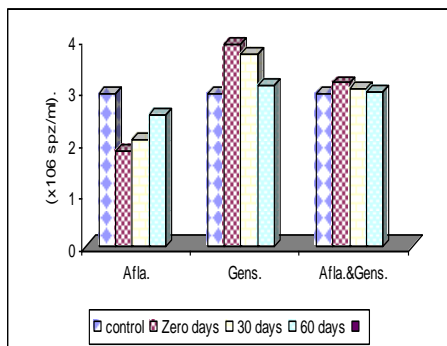
** Significant at P<0.01

Table 4: Effect of aflatoxin (1 mg /kg BW) gensing (200 mg /kg BW) either alone or together on some liver and kidney functions at 1st, 30th and 60th days post medication for two months in male baladi rabbits (n=5).

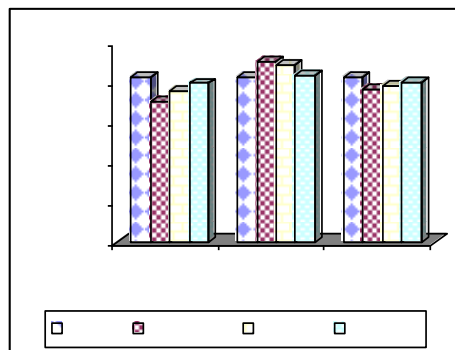
parameter	Healthy rabbit	1 st days			30 th days			60 th days		
		AF	GE	AF+GE	AF	GE	AF+GE	AF	GE	AF+GE
AST (U/L)	41.36± 1.76	50.62± 2.46*	42.52± 1.54	40.35± 1.83	48.62± 1.54*	41.83± 1.05	40.73± 2.62	40.62± 1.63	41.68± 1.48	41.51± 1.89
ALT (U/L)	17.51± 1.83	25.93± 1.92*	18.34± 0.68	16.03± 0.93	23.83± 1.06*	17.92± 1.69	16.82± 1.63	19.65± 1.62	17.54± 0.93	17.48± 0.72
Alk.ph. (I.U/ml)	63.12± 2.15	70.36± 1.83*	63.61± 2.08	62.93± 1.82	68.93± 1.71*	62.91± 2.94	63.04± 1.39	65.93± 1.48	63.06± 1.63	63.19± 1.94
T. protein (gm/dl)	7.61± 0.59	5.21± 0.33**	10.43± 0.84*	7.20± 0.57	5.97± 0.23*	9.57± 0.41*	7.27± 0.76	7.30± 0.44	7.93± 0.40	7.60± 0.60

*Significant at P < 0.05

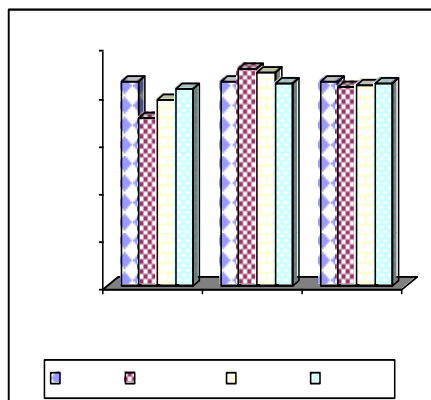
** Significant at P<0.01



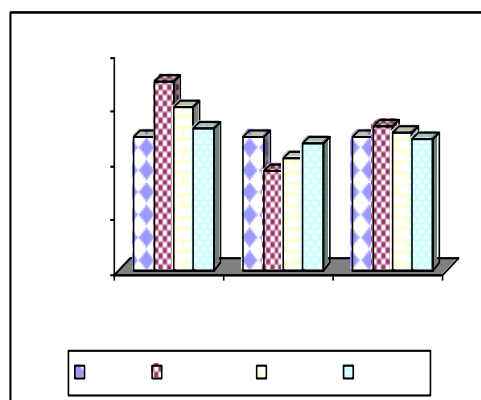
A) Sperm cell concentration



B) Sperm motility



C) Alive sperm



D) Total sperm abnormality

DISCUSSION

The obtained results revealed that aflatoxin caused highly significant decrease in the sperm cell concentration, sperm motility, alive sperm and induced significant increase in sperm abnormalities at 1st and 30th days post the use of aflatoxin. This effect was in accordance with that obtained by Baker and Greene (1987) who found that aflatoxin induced significant decrease in sperm cell concentration, progressive motility and alive percent of spermatozoa from 91.11 % to 80.8%. Another explanation were reported by Sahoo, *et al.* (1993) Who recorded that the decreased in sperm cell concentration, progressissive motility may be attributed to the disruption of spermatogenesis by chronic aflatoxicosis which is preceded and caused by the impairment of the leydig cell functions and the resultant drop in testosterone in the testis in rats. Ragheb and Sahar (2003) mentioned that the fertility of

animals was vigorously reduced by aflatoxin consumption due to reduce in LH and FSH in male rats which are responsible for regulation of normal productivity of rats.

Significant decrease in the mean values of blood serum testosterone at 1st and 30th days post addition of aflatoxin for 60 days in the present study. Close similarity was seen between the finding and those obtained by Hassan *et al.* (2004) who found that aflatoxin induced decrease in testosterone hormone due to occurrence of oedema, fibrosis and local leydig cell.

Gensing treatment was able to increase the sperm cell concentration, sperm motility, alive sperm and testosterone hormone and induced significant decrease in sperm abnormalities at 1st and 30th days post treatment. The previous finding fit in with those previously reported by El-Sayed (2005) in rabbit. Moreover, this is supported by the findings of Yamamoto, *et al.* (1977) who said that gensing stimulated spermatogenesis in rat and rabbit testes and attributed to the stimulatory effect of ginsenosides on spermatogenesis to stimulation of DNA and protein synthesis in testis (Salvati *et al.* 1996). Ginsenosides may have an effect at different levels of the hypothalamus, pituitary, testis axis as increases hypothalamic GnRH, pituitary gonadotropins and testosterone levels in the blood in male rats (Fahim *et al.*, 1982). Keeping with this line, Mkrtchyan *et al.* (2005) observed that gensing resulted in an increase in the number of spermatozoa, normal sperm motility and fertility indexes. The improvement in number of spermatozoa and motility in the present study may be due to the activation of spermatogenesis with gonadotropins hormones secreted from pituitary gland under the effect of GnRH (Nasr *et al.*, 1994). This is supported by the findings of Sung *et al.*, (2000) who said that Korean red gensing can improve the vascular endothelial dysfunction in patients with hypertension possibly through increasing NO. NO is a mediator that cause vascular smooth muscle fibers to relax thus resulting in vasodilatation, increasing the blood flow thus can improve the function of sexual organs (Huang, 1993). Decrease in sperm abnormalities in the present investigation may be due to the fact that androgen is essential for most stages of spermatogenesis and sperm production, furthermore, vitamin B12 (one component of gensing) participates in the synthesis of DNA, which conjugated with histon to form nucleoprotein of the sperm nucleus (Felsenfeld, 1978).

The present study illustrated that rabbits treated with gensing revealed significant increase in testosterone at 1st, 30th and 60th days post

medication. Close similarity was seen between the finding and those obtained by El-Sayed, (2005) who found that chronic administration of gensing produced a dose related increase in testosterone levels in male rats. This ultimately results in stimulation of steroidogenesis and production of testosterone (Hafez, 1987). In the current work, it has been found that aflatoxin induced significant decrease in weight of testis, seminal vesicle and prostate gland. The above mentioned results were supported by previous studies of Alexander (1978). The previous author mentioned that the development and maintenance of accessory sex organs and their secretion depend on androgen, Based on this idea the significant decrease in weight of accessory sex organs of rabbit feed on ration containing aflatoxin could be due to decrease in testosterone hormone level which recorded in the present study. Administration of gensing to rabbit induced non significant change in the sex organs weight in comparison to control group and these results run parallel with those obtained by Murphy *et al.* (1998) and El-sayed, (2005).

The present investigation revealed that the aminotransferases (AST-ALT) and alkaline phosphatase were significantly increased at 1st and 30th days post feeding rabbit on ration contain aflatoxin. Our results came in agreement with Arafa, *et al.* (2006). Mehta, *et al.* (1993) who stated that, aflatoxin induced hepatotoxicity and revealed a significant increase in the activity of plasma AST and ALT due to leakage of these enzymes into circulation in male rats. Another explanation for the increased activities of the liver enzyme occurs due to altered permeability of hepatocytes (Roger *et al.*, 1991).

Results of the present study revealed non significant effects on aminotransferases (AST- ALT) and alkaline phosphatase concentrations at 1st and 30th days post administration of gensing in rabbits. These results agree with those of Lin, *et al.* (1995). They recorded that liver functions were unchanged after the use gensing extract in rats.

On the other context co-treatment of gensing and aflatoxin improve these adverse effects on the semen picture, testosterone and liver functions in comparison to control group. Rabbits treated with gensing and aflatoxin resulted in significant effect when compared with that of aflatoxin treated rabbit. This result can be explained by that reported by El-Saieed, (2003) who stated that, gensing exerted antioxidant effect by enhance the activity of the antioxidant enzymes. In addition, it has the ability to directly neutralize a number of toxic reactants and stimulate antioxidative enzymes. Also gensing inhibited

the hepatic damage induced by toxin, Gensing also normalized liver functions and the serum liver enzyme activities (AST, ALT, Alkaline phosphatase) were not differ in compared with control group. These present finding suggested that gensing could protect hepatocytes membrane against oxidative stress induced by aflatoxin and hence prevent the libration of liver enzymes to the sera so it support liver functions. Okada and Zhang (1998) reported that gensing could protect cell membrane fatty acids from decomposition induced by free radicals so support the all functions of the animal body. Gensing also has a diuretic effect, produce light diarrhoea lead to decrease the amount of absorbed toxin and increase the level of glutathione (Reylnoids, 1991). Another suggestion is that gensing may repair the resources of some antioxidant enzymes as it increase the level of glutathione (El-Saieed, 2003).

In conclusion, from the present results it appears that aflatoxicosis in rabbits affects the male fertility and biochemical parameters. Therefore it is much better to control aflatoxicosis by using healthy feed free from aflatoxin and clean environ-ment rather than treatment of the rabbits.

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