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**BIOCHEMICAL AND PATHOLOGICAL STUDIES OF
COBALT LEVEL IN SLAUGHTERED CATTLE AND
BUFFALOES IN ASSIUT GOVERNORATE**
(With 3 Tables and 7 Figures)

By

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**دراسات بيوكيميائية وباثولوجية عن مستوى الكوبالت في الأبقار والجاموس
المذبوحة في محافظة أسيوط**

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أجريت هذه الدراسة للتعرف على القيم المختلفة لمستوى الكوبالت في الأبقار والجاموس قيد الدراسة والتغيرات البيوكيميائية والباثولوجية المصاحبة لنقص مستوى الكوبالت. حيث ثبت هذا النقص بعد تقدير مستوى الكوبالت في مكونات مصل الدم المختلفة وايضا في نسيجي الكبد والكلى. تم إجراء الدراسة على عدد ٥٠ حيوان منهم ٢٥ من الأبقار و ٢٥ من الجاموس. تم تجميع عينات الدراسة بصورة عشوائية لإستخدامها في إجراء الفحص الكيميائي والباثولوجي. وقد استخدمت عينات (الكلى والكبد) للفحص الهستوباثولوجي. تم في هذه الدراسة مناقشة التغيرات البيوكيميائية والباثولوجية الناتجة عن نقص مستوى الكوبالت المنعكس على صورة الدم وكذلك مكونات مصل الدم وايضا في نسيجي الكبد والكلى. كما تأكد هذا النقص ايضا بعد فحص مستوى الكوبالت في الماء والغذاء المقدم لهذه الحيوانات في داخل المزرعة التي تم تربيتهم بها. وقد أظهر الفحص الباثولوجي للحيوانات التي كانت تعاني من نقص في مستوى الكوبالت من التقرز في الخلايا الكبدية مع وجود إحلال دهني للأنسجة الكبدية في بعض المناطق. كما وجد في بعض الحالات تكاثر الأنسجة الضامة المحيطة بالاوعية المرارية مع إنتشار الخلايا الليمفاوية بها. أما الكلى فقد كانت تعاني من انحلال الخلايا الطلائية المبطنة لأنابيب الكلى مع وجود جلطات متفرقة في الكبيبات.

SUMMARY

Fifty animals, cattle and buffaloes (25 each) aged from 5-7 years old were selected randomly. They were raised at governmental project and slaughtered in Bani Adi and Elhawatka slaughter houses (Assiut, Egypt). Random samples of feed stuff and water (10 for each) were taken from the farm where the animals were raised on. Haemogram showed that cattle and buffaloes suffering from cobalt deficiency had significantly decreased values of RBCs, PCV, Hb, MCV, MCH, MCHC and WBCs. According to serum biochemistry: cattle and buffaloes had lower levels of cobalt, significantly lower values of total proteins, albumin, and globulin. Moreover, the activities of AST and ALT and the concentrations of blood urea nitrogen showing a marked increase in the animals suffering from lower cobalt levels. According to the histopathological findings: Livers of cobalt deficiency animals were exhibited varying degrees of vacuolar degeneration and necrotic changes with cytoplasmolysis and karyorrhexis of the hepatic nucleus reached to its lyses. Diffuse fatty changes were also observed. The central veins were congested and surrounded with degenerated hepatocytes and increased number of kupffer cells. Some cases showed portal cirrhosis with lymphocytic infiltration. The kidneys showed severe degenerative changes in the renal tubules including cytoplasmolysis in the cytoplasm and lyses of the nucleus, congestion of the glomeruli were also observed. Therefore we should follow up cobalt levels in our farm animals to prevent its deficiency in our farms to prevent its deficiency in our farms in Assiut Governorate.

Key words: Cobalt, cattle, buffaloes, haemogram, histopathology.

INTRODUCTION

Cobalt (Co) is an essential trace element in ruminant diets for the production of vitamin B12 by the rumen microbes to meet the vitamin B12 requirements of both the ruminal bacteria and the host animal (McDowell, 1992). In higher animals, vitamin B12 is a cofactor for two enzymes, methylmalonyl-CoA mutase and methionine synthase.

The former catalyzes the interconversion of methylmalonyl-CoA to succinyl-CoA (Banerjee and Chowdhury, 1999), an important step in gluconeogenesis, while the latter acts to remethylate homocysteine, in the terminal step of methionine synthesis (Matthews, 1999). The NRC (2001) lists the dietary requirement of dairy cattle for Co as 0.11 mg/kg

DM; relatively little is known about Co metabolism in cattle. Ruminants fed forages with Co concentrations <0.08 mg/kg develop signs of Co/vitamin B12 deficiency (McDowell, 1997).

Necropsy of severely affected animals shows emaciation, often with total absence of body fat, liver fatty changes and occasional spleen haemosiderosis (Paterson and MacPherson, 1990; Kennedy *et al.*, 1994) lambs are the most sensitive to Co deficiency, followed by mature sheep, calves, and mature cattle (McDowell, 2003; McDowell and Arthington, 2005).

Ruminants appear to be more sensitive to vitamin B12 deficiency than non-ruminants (NRC, 2001).

Elemental Co is absorbed through the intestinal tract and transported in blood to various tissues. The liver and the kidney contain the highest concentration of Co within tissues and are considered the main storage site (Underwood and Suttle, 1999).

Besides covering the requirements for vitamin B12 synthesis, Co may play a role in rumen fermentation by increasing fiber digestion from low quality forages (Zelenak *et al.*, 1992; Stemme *et al.*, 2008; Girard *et al.*, 2009; Kumar *et al.*, 2011).

In the present study, analysis of the animal feed, water, blood, blood serum, liver and kidney was conducted to establish the different Co levels in some selected individuals representing cattle and buffaloes and its effect in the different parameters in this study.

MATERIALS and METHODS

Animals:

Fifty animals, cattle and buffaloes (25 each) aged from 5-7 years old were selected randomly. They were chosen at governmental project and slaughtered in Bani Adi and Elhawatka slaughter houses (Assiut, Egypt).

Blood sampling:

Jugular blood samples were collected on EDTA for haematological evaluation and in plain vacutainer tubes for serum biochemistry. The blood was allowed to clot and centrifuged, then clear blood serum was separated and stored at -20 C° until analysed.

Haematological parameters: Red blood cell counts (RBCs), packed cell volume (PCV), haemoglobin (Hb) and white blood cell counts (WBCs) were determined according to Feldman *et al.* (2000). Red blood

cell indices; mean corpuscular volume (MCV), mean corpuscular haemoglobin (MCH) and mean corpuscular haemoglobin concentration (MCHC) were calculated.

Serum biochemistry: blood serum were analyzed spectrophotometrically for determination of total proteins (Henary *et al.*, 1974), albumin (Doumas *et al.*, 1971), and transferases (Aspartate amino transferase; AST and alanine amino transferase; ALT), and blood urea nitrogen (Patton and Crouch, 1977) using test kits supplied by Stanbio – ITALY.

Collection of liver and kidney samples:

Liver and kidney samples were collected from the selected slaughtered cattle and buffaloes. The samples were transferred into clean sterile containers and immediately frozen at -20 °C until analysed.

3- Pathological studies:

Tissue specimens from liver and kidneys were obtained immediately from the freshly slaughtered animals and fixed in 10% neutral formalin solution dehydrated, cleared and embedded in paraffin blocks. Paraffin sections of 5 μ thickness were prepared, stained by haematoxylin and eosin (H&E) and osmium tetroxide for demonstration of fat according to Bancroft *et al.* (1996). These stained sections examined microscopically for detection of histopathological alterations.

Food samples: Ten samples of concentrates and hay were taken and analysed using an atomic absorption spectrophotometer. Two grams of each sample was wet ashed in a Teflon beaker with cover using (1:3) HNO₃/HClO₄ acid mixture. The residue after evaporation was dissolved in dilute HCL and completed to 50 ml using bi-distilled water according to the method of Henary *et al.* (1974).

Biochemical analysis:

Blood serum concentrations of cobalt were measured by atomic absorption spectrophotometer (B3003, Perkin Elmer-AAS). Liver and kidney samples (one gram) were digested in a mixture of 2:1: 0.5 nitric acid (HNO₃, 65%, Perchloric acid (HClO₄, 60%) and sulphuric acid (H₂SO₄, 97%). The samples were further diluted and aspirated into an atomic absorption spectrophotometer.

Statistical analysis: Recorded data were analyzed statistically using analysis of variance (ANOVA). The statistical differences between

means were estimated by Duncons Multiple Range test. The computation was facilitated by statistical package SPSS (2000).

RESULTS

According to blood serum cobalt analysis, cattle (0.23 ± 0.02 $\mu\text{g/ml}$) and buffaloes ($0.25\pm 0.05\mu\text{g/ml}$) (Table 1) were considered Co-deficient animals when comparing with the adequate levels of serum cobalt (0.28 ± 0.05 $\mu\text{g/ml}$) recorded by Kincaid *et al.* (2003) and need more care for Co supplement.

In this study The cobalt level of the liver (Table 1) in cattle was 0.06 ± 0.02 mg/Kg and in buffaloes was 0.08 ± 0.03 mg/Kg. while the kidney Co levels in cattle and buffaloes were (0.03 ± 0.01 mg/Kg) and (0.05 ± 0.02 mg/Kg) respectively.

Liver and kidney of large ruminants should be >1.00 $\mu\text{mol/kg}$ DM in the normal conditions which consider an adequate amount of Cobalt as reported by Tiffany *et al.* (2003).

Our study revealed that the level of cobalt in forages and water (Table 1) introduced to the studied animals were 0.023 p.p.m and 0.01 p.p.m in cattle and 0.030 p.p.m and 0.01 p.p.m in buffaloes respectively. The levels of cobalt in forage and water considered decreased when it is <0.10 mg/kg DM as recorded by Larry (2005)

Haemogram: cattle suffering from cobalt deficiency had significantly decreased ($P<0.05$) values (Table 2), RBCs (12.41 ± 0.63 T/l), PCV ($31.88\pm 0.63\%$), Hb (9.16 ± 0.25 g/dl.), MCV (28.02 ± 2.65 fl), MCH (7.63 ± 0.30 pg), MCHC (31.83 ± 2.10 g/dl and WBCs (7.49 ± 0.10 ($10^3\mu\text{l/l}$)). Also buffaloes suffering from cobalt deficiency had significant decreased ($P<0.05$) values (Table 2), RBCs (11.03 ± 0.30 T/l), PCV($32.3\pm 1.36\%$), Hb (8.6 ± 0.48 g/dl), MCV(28.08 ± 2.85 fl), MCH (7.79 ± 0.30 pg), MCHC (30.88 ± 2.0 g/dl) and WBCs (8.64 ± 0.12 ($10^3\mu\text{l/l}$)) than the normal values of RBCs (13.43 ± 0.34 T/l), PCV($40.45\pm 0.62\%$), Hb (13.56 ± 0.27 g/dl), MCV (29.05 ± 2.65 fl), MCH (9.77 ± 0.34 pg), MCHC (33.42 ± 0.29 g/dl) and WBCs (11.81 ± 0.15 ($10^3\mu\text{l/l}$)) reported by (Stangl *et al.*, 1999).

Serum biochemistry: Cattle and buffaloes suffering from cobalt deficiency had significant lower ($P<0.05$) values (Table 3) of total proteins (4.01 ± 0.06 and 5.04 ± 0.07 g/dl), albumin (1.67 ± 0.07 and 1.78 ± 0.04 g/dl), and globulin (2.54 ± 0.10 and 2.86 ± 0.17 g/dl) respectively, than the normal levels reported by Singh and Chhabara

(1995) where the normal serum total proteins (6.88 ± 0.28 g/dl), albumin (2.60 ± 0.12 g/dl), and globulin (4.26 ± 0.24 g/dl).

The activities of serum enzymes (Table 3) were (244.23 ± 11.8 and 327.34 ± 14.7 IU/l for AST in cattle and buffaloes respectively) and (38.12 ± 1.09 and 40.0 ± 1.34 IU/l) for ALT in cattle and buffaloes respectively. The concentrations of blood urea nitrogen was (15.11 ± 1.07 and 16.11 ± 1.11 mg/dl) in cattle and buffaloes respectively, showing a marked increase in the lower cobalt animals. Normal values of the activities of AST was (129.42 ± 9.11 IU/l) and ALT was (17.15 ± 0.69) and the normal concentration of blood urea nitrogen was (12.21 ± 0.31) recorded by Singh and Chhabara (1995).

Table 1: Cobalt levels in Serum, liver, kidney, forage, water of cattle and buffaloes selected in this study:

	Cattle	Buffaloes
Serum $\mu\text{g/ml}$	0.23 ± 0.02	0.25 ± 0.05
Liver mg/Kg	0.06 ± 0.02	0.08 ± 0.03
Kidney mg/Kg	0.03 ± 0.01	0.05 ± 0.02
Forage DM basis p.p.m	0.023	0.030
Water p.p.m	0.01	0.01

Table 2: Haemogram of the selected cattle and buffaloes for this study:

	Cattle	Buffaloes
Red blood corpuscles(RBCs,T/l)	12.41 ± 0.63 T/l	11.03 ± 0.30
Packed cell volume (PCV %)	31.88 ± 0.63	32.3 ± 1.36
Haemoglobin (Hb-g/dl)	9.16 ± 0.25	8.6 ± 0.48
Mean corpuscular volume (MCV-fl)	28.05 ± 2.65	29.06 ± 2.85
Mean corpuscular haemoglobin (MCH-pg)	7.63 ± 0.30	7.79 ± 0.30
Mean corpuscular haemoglobin concentration (MCHC-g/dl)	31.83 ± 2.10	30.88 ± 2.0
Total leukocytic count (WBCs $\times 10^3 \mu\text{l/l}$)	7.49 ± 0.10	8.64 ± 0.12

Mean \pm SE. SE= standard Errors. *=significant at $p=0.05$.

Table 3: Serum biochemical parameters of the selected cattle and buffaloes for this study:

	Cattle	Buffaloes
Total proteins(g/dl)	4.01±0.06	5.04±0.07
Albumin(g/dl)	1.67±0.07	1.78±0.04
Globulin(g/dl)	2.54±0.10	2.86±0.17
Albumin globulin ratio	0.69±0.05	0.72±0.07
Aspartate amino transferase(AST-IU/l)	244.23±11.8	327.34±14.7
Alanine amino transferase(ALT-IU/l)	38.12±1.09	40.0±1.34
Urea nitrogen(mg/dl)	15.11±1.07	16.11±1.11

Mean ±SE. SE= standard Errors. *=significant at p=0.05.

Gross examination:

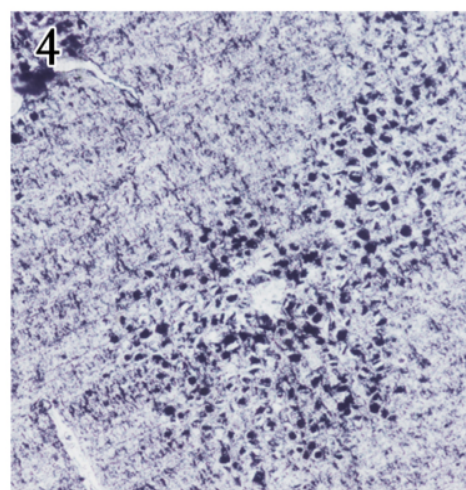
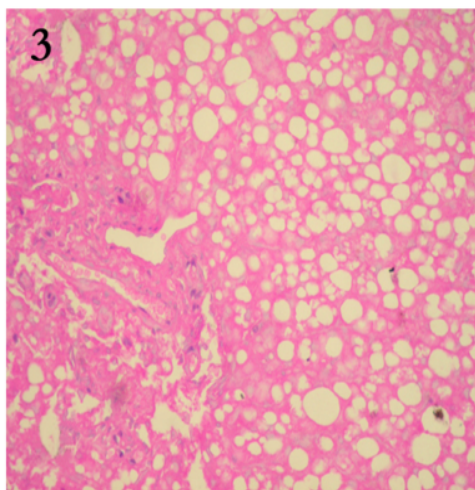
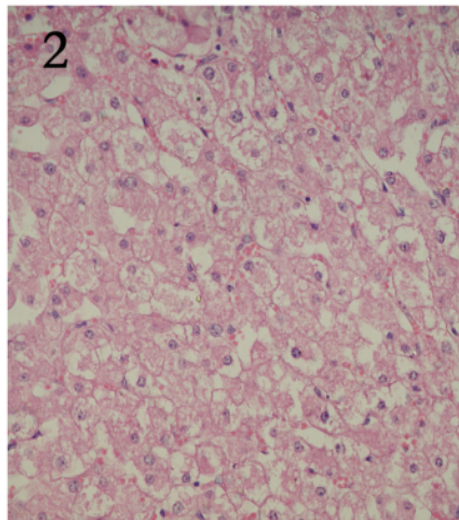
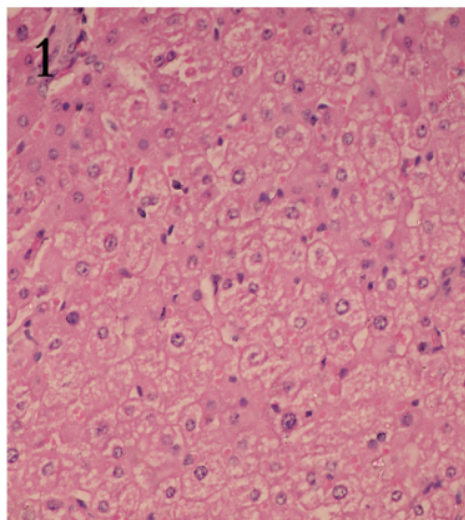
The livers were characterized as uniformly pale, swollen and distinctly friable. It's cut surface is greasy and yellowish in color.

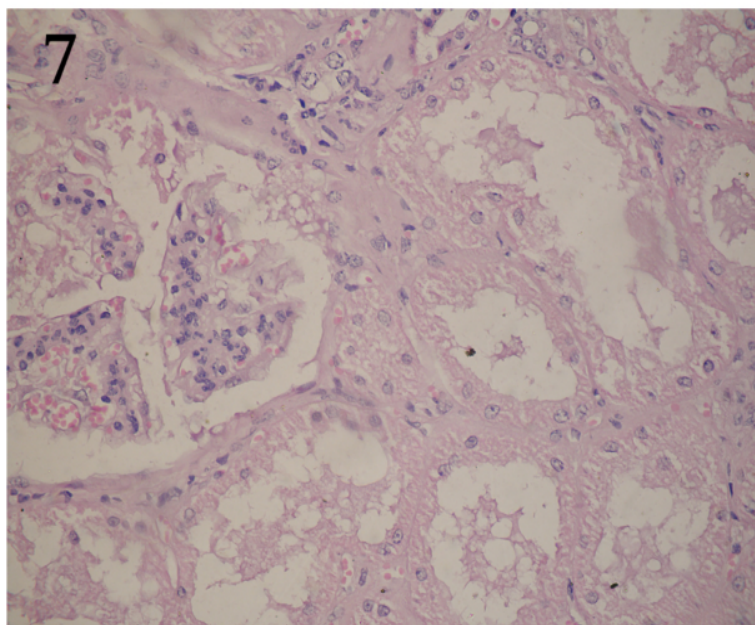
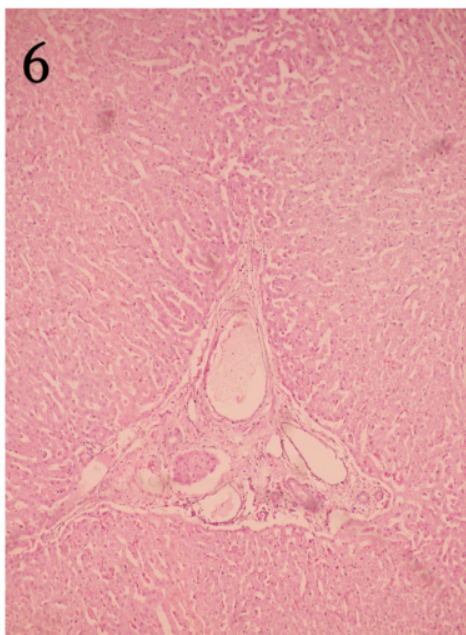
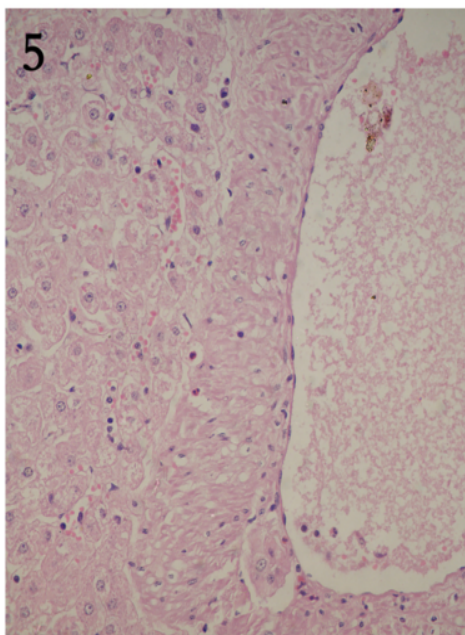
Histopathological examination:

Livers of Co-deficiency animals were exhibited varying degrees of vacuolar degeneration and necrotic changes with cytoplasmolysis and karyorhxis of the hepatic nucleus reached to its lyses (Fig 1,2). Diffuse fatty changes was also observed (Fig 3,4). The central veins were congested and surrounded with degenerated hepatocytes and increased number of kupffer cells (Fig 5) some cases showed portal cirrhosis with lymphocytic infiltration (Fig 6). The kidneys showed sever degenerative changes in the renal tubules including cytoplasmolysis in the cytoplasm and lyses of the nucleus congestion of the glomeruli was also observed (Fig 7).

LEGEND OF FIGURE

- Fig. 1:** Liver of cobalt deficient animal showed vacuolar degeneration of the hepatocytes H&E X40.
- Fig. 2:** Liver of cobalt deficient animal showed degenerative changes in the hepatocytes including kayorhexis of the nucleus and lyses of the cytoplasm hepatocytes H&E X40.
- Fig. 3:** Liver of cobalt deficient animal showed fatty degeneration H&E X10.
- Fig. 4:** Liver of cobalt deficient animal showed fatty degeneration osmium tetraoxid stain X10.
- Fig. 5:** Liver of cobalt deficient animal showed congestion of the central vein which surrounded with degenerated hepatocytes and increased number of kupffer cells H&E X40.
- Fig. 6:** Liver of cobalt deficient animal showed portal cirrhosis with infiltration of the portal area with lymphocytes H&E X10.
- Fig. 7:** Kidney of cobalt deficient animal showed degenerative changes in both renal tubule and the glomeruli H&E X40.





DISCUSSION

In general, cobalt deficiencies are frequent when high levels of mixed concentrates are fed. Cobalt deficiency in ruminants is of a major economic importance in several countries (Kennedy *et al.*, 1997). More recent studies indicate the necessity to increase the amount of dietary Co for growing ruminants up to a level of 300-500 µg/Kg DM for optimum microbial activity, fermentation and vitB12 synthesis (Paragon, 1993 and Singh, Chhabara, 1995; Anonymous, 1996). Dietary Co requirements have been established at 0.1 to 0.3 ppm (NRC, 2001). Signs of Co deficiency develop in ruminants fed diets that contain less than 70 µg/Kg DM (Marston, 1970; Lopez-Guisa and Satter, 1992 and Tiffany, 2003).

The present study showed that cobalt level in (blood serum, blood, liver, kidney, forages, and water) are considerably deficient for both cattle and buffaloes, in comparing with the different adequate cobalt levels recorded by Singh and Chhabara (1995); Stangl *et al.* (1999); Kincaid *et al.* (2003); Tiffany *et al.* (2003); Larry (2005).

Similar findings were reported by Khan (2003); Khan *et al.* (2006 and 2008).

In the current study, there was significant decrease in erythrogram parameters, (RBCs, PCV, Hb, MCV, MCH and MCHC) in Co deficient animals (cattle and buffaloes), in comparing with normal healthy values reported by other researches (Stangl *et al.*, 2000; Khan *et al.*, 2006 and 2008).

The Co deficient animals showed mild microcytic hypochromic anemia as manifested by reduced MCV and MCHC.

The prominent feature of this study might be the decreased RBCs and Hb, in these animals. These results give an indication about the importance of Co and its essential role in producing vit B12 and folate for the production of haem, Stangl *et al.* (1999).

The obtained results showed that the Co-deficient cattle and buffaloes exhibited a significant decrease in total serum proteins, albumin and globulin. Impairment of protein synthesis may be the principal reason for growth depression frequently observed in these animals. It may be also due to lower digestibility coefficients for dry matter (Singh and Chhabara, 1995; Stangl *et al.*, 1999).

Also in this study Co, depleted cattle and buffaloes had elevated serum activities of AST and ALT., these findings are indicative of

primary hepato cyte damage. Similar results were obtained by (Kennedy *et al.*, 1997).

The histopathological findings as reported in this study support primary hepatocyte damage as the main lesion in liver of Co- deficiency animals. Although AST is not a liver specific enzyme, it has been reported to be elevated in cattle (Cebra *et al.*, 1997) with hepatic lipidosis.

Significant elevation of blood urea nitrogen, as observed in the present study has also reported by (Stangl *et al.*, 1999) in Co- deficient cattle and buffaloes. This result may be attributed to the disturbance in protein metabolism due to liver damage, which leads to accumulation of urea in blood (Kaneko, 1989).

Keeping in view this deficiency of cobalt in forages, water and intern in animals, best be prevented by direct oral intake of Co through mineral supplements (Khan *et al.*, 2006 and 2008).

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