

MICROBIAL HAZARDS ASSESSMENT DURING WHITE CHEESE PRODUCTION IN A TRADITIONAL DAIRY PLANT

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ABSTRACT

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This study has been conducted to determine the microbiological contamination sources during production of white soft cheese in a local dairy plant, in Alexandria city, Egypt. A total of 180 representative samples (coming raw milk, cheese vat, cheese mould, wooden tables, cheese handler ,water supply, wall, plastic package and white soft cheese) 20 of each. The trials repeated 3 times to ensure the most contaminated points. The highest total mesophilic bacterial count was found in Cheese mould with $9.4 \times 10^6 \pm 3.7 \times 10^4$ cfu/cm² followed by Plastic package with $2.1 \times 10^6 \pm 3.6 \times 10^4$ cfu/ cm². The most contaminated point with *coliforms* was cheese vat with a mean count of $6.1 \times 10^4 \pm 1.11 \times 10^3$ followed by cheese handler with a mean count of $5.6 \times 10^3 \pm 1.8 \times 10^2$. The most contaminated point with a total psychrotrophic count was cheese vat with a mean count of $4.5 \times 10^5 \pm 6.4 \times 10^4$ followed by water supply with a mean count of $3.7 \times 10^5 \pm 1.9 \times 10^4$. The highest *S aureus* count was found in cheese handler by $9.4 \times 10^4 \pm 5.6 \times 10^3$ followed by the wall with a mean count of $8.5 \times 10^3 \pm 2.3 \times 10^3$. *Enterococci* count contaminated wooden table with a mean value of $2.8 \times 10^4 \pm 1.3 \times 10^2$ followed by water supply with a mean count of $1.3 \times 10^3 \pm 4.7 \times 10^2$. The most contaminated point with yeast and mould was cheese mould with a mean count of $5.8 \times 10^3 \pm 1.3 \times 10^2$ followed by wooden table with a mean count of $3.4 \times 10^3 \pm 1.6 \times 10^2$. The most contaminated points which affect the microbial quality of white soft cheese during processing were cheese mould followed by wooden table, cheese handler and cheese vat. The application of the hygienic measures for manufacturing of the dairy products greatly improves their quality. The hygienic measures include cooling of the milk, heat treatment, use of clean water, personal hygiene of cheese handler, sterilization of equipment and prevent of post contamination during manufacture. A HACCP – based risk assessment and good manufacturing practice should be employed for all stages of manufacture of soft cheese.

Key words: HACCP, White cheese, Dairy plant

INTRODUCTION

The main potential hazards in most dairy products are microbiological hazards and the dairy industry has increased its efforts for quality and safety assurance through the development and implementation of protective programs as HACCP and quality assurance system. (Kassem *et al.*, 2002). White soft cheese is one of the common delicious cheeses consumed in Egypt. Their manufacture and handling techniques in Egyptian markets are still primitive and unhygienic, Many contaminants find their way to raw milk, from which they gain access to dairy products (Sadek *et al.*, 2009).

The successful commercial production of dairy products requires control of microbiological presence and activity to achieve maximum shelf-life consistent with safety of the product (pelecszynska and Libelt, 1995).

The microbial quality and safety of Egyptian white soft cheese, is the major area of concern for

producers, public health authorities and consumers. It depends on the types of microorganisms introduced from raw milk, efficiency of processing and the hygienic practice applied in small or big dairy plant or informal producers. Handling of milks during cheese manufacture play an important role in the proliferation of microbial flora and consequently impair its utility and render the product unfit for human consumption (Aly and Galal., 2002). The microbial quality of used raw milk in soft cheese has an important role in the quality of yielded cheese. Many researchers have recorded unlimited microbial flora in the raw milk (Hayes *et al.*, 2001).

Food borne diseases are a common and widespread global problem. Several outbreaks have been reported as a result of eating contaminated dairy food that may look, taste and smell perfectly normal but are in fact contaminated with large number of harmful bacteria (CDC, 2009).

Egyptian standard (2005) stated that soft cheese must be free from any pathogenic microorganisms, while

coliform counts must be less than 10 cfu/g, total yeast counts less than 400 cfu/g and total mold counts less than 10 cfu/g.

Microorganisms may gain access to cheese during process, handling and distribution since milk provide a high nutritive, favorable media for the growth and multiplication of such organisms. Many food poisoning outbreaks may be due to using milk from diseased animals with infection of bacterial origin or manufacturing in contaminated places or from the workers themselves. Ingestion of certain microorganism can be detrimental to human health (UNEP, 1992). Coliforms are routinely used as indicator to a certain the quality of the food products. Their presence indicates careless methods of production, handling of the processed food products and the use of insufficient sanitized equipment. Moreover, Coliforms are used to measure the quality of the practices used to minimize microbial contamination of dairy products and as an approved safety indicator in HACCP system (Banwart, 1998).

The presence of *staphylococcus aureus* in dairy products is a good indicator of the personal hygiene of factory workers, high *coliform* in these products induce undesirable changes that lower their quality, presence of moulds and yeasts in dairy products is undesirable even when found in few numbers as they rapidly grow in the product that render it of inferior quality (Hamed, 1992).

Handling of milk during cheese manufacture plays an important role in the proliferation of microbial flora and consequently impairs its utility and renders the product unfit for human consumption (Yousef *et al.*, 2001). Mould and yeast counts in cheese are used as an index of the proper sanitation and quality defects in soft cheese as rancidity, softness and colour defects arise mainly from contamination by yeast and mould. Moreover, in view of the potential ability of some mould to produce mycotoxins during their growth thus, their presence posse potential hazards to food safety and human illness (Besancon *et al.*, 1992).

Hazards of the dairy products mainly microbiological hazard and can provide a toll for evaluating the acceptability of the products or process designed to control presence or growth of microorganisms (Silliker, 1987).

Microbiological investigations for the different varieties of Egyptian white soft cheese have been carried out either to evaluate their qualities, hinder or minimize microbial spoilage and to determine the cheese safety as free from food borne microorganisms, (Abou-Dawood *et al.*, 2005).

The aim of this work to assess the potential microbial Hazards in traditional dairy plant in Alexandria city

through the evaluation of its microbial criteria and giving suitable recommendations to improve the hygienic quality of the dairy plant to prevent contamination and produce safe and high quality products.

MATERIALS and METHODS

The present study was carried out in a traditional dairy plant in Alexandria city.

1-Process flow diagram of soft cheese:-

Incoming raw milk →cheese vat →addition of salt (8.5- 9.5%) →pouring of milk→ heating to 38 c° →Renneting vats→ Addition of rennet→ wooden moulds→ wooden cheese table →pressing→ Blocking →Wrapping in polyethylene paper bags →End product (soft cheese).

2- Collection of samples:-

2-1- Collection and preparation of samples: 180 samples were taken from different sites of dairy processing lines from reception to packaging. Representative as (incoming milk- cheese vat- cheese mould- wooden table- - cheese handler-water supply-wall- plastic package- soft cheese) 20 of each. The trials repeated 3 times to ensure the most contaminated point.

2-2- Incoming milk: the samples was collected in sterile sampling bottles after through mixing of the bulk(about 500 ml each),milk in cheese vat about 500 ml of milk were collected after pouring into cheese vat in a sterile screw capped bottle.

2-3-Soft cheese samples: the samples was collected in sterile sampling bottles (250gm) each sample thoroughly mashed in a sterile mortar for microbiological examination. The samples were collected under hygienic condition and dispatched directly to the laboratory with a minimum of delay in an ice- box (at 4 ± 1 c°).

2-4-Other samples: samples from (cheese vat, wooden cheese table, cheese moulds and workers' hands) were collected by swab contact method (Pritchard *et al.*, 1994). Samples from plastic packages were collected by Rinse solution method and water supply samples (A.P.H.A., 1992).

3- Microbiological examination:-

3-1-Total mesophilic bacterial count (ISO4833/2003).

3-2- *Coliforms* count (ISO 1991).

3-3- Total psychrotrophic count (APHA, 1992).

3-4- *Staphylococcus aureus* count (FDA, 2002).

3-5- Total Yasts and Moulds count (Bailey and Scott, 1998).

3-6-Total *Enterococci* count (Efthymiou *et al.*, 1974).

RESULTS

Table 1: Statistical analytical results of Total mesophilic bacterial count and Coliforms count during processing steps of white soft cheese in traditional dairy plant.

Item	total mesophilic bacterial count	coliforms count
Incoming raw milk	$3.2 \times 10^3 \pm 1.3 \times 10^4$	$4.2 \times 10^3 \pm 7.2 \times 10^2$
Cheese vat	$4.6 \times 10^5 \pm 1.9 \times 10^4$	$6.1 \times 10^4 \pm 1.11 \times 10^3$
Cheese mould	$9.4 \times 10^6 \pm 3.7 \times 10^4$	$2.1 \times 10^2 \pm 0.28 \times 10$
Wooden table	$8.5 \times 10^5 \pm 4.6 \times 10^3$	$6.2 \times 10^2 \pm 3.5 \times 10$
Cheese handler	$7.5 \times 10^5 \pm 6 \times 10^4$	$5.6 \times 10^3 \pm 1.8 \times 10^2$
Water supply	$9.4 \times 10^5 \pm 1.7 \times 10^3$	$3.9 \times 10^2 \pm 1.6 \times 10^2$
Wall	$5.8 \times 10^5 \pm 4.8 \times 10^4$	$5.7 \times 10^2 \pm 4.3 \times 10^2$
Plastic package	$2.1 \times 10^6 \pm 3.6 \times 10^4$	$3.8 \times 10^2 \pm 1.4 \times 10^2$
Soft cheese	$3.9 \times 10^7 \pm 2.8 \times 10^5$	$2.4 \times 10^2 \pm 1.3 \times 10$

The highest total bacterial mesophilic count was found in cheese mould followed by plastic package. The most contaminated point with coliforms was in cheese vat followed by cheese handler.

Table 2: Total psychrotrophic count and *S. aureus* count during processing steps of white soft cheese in traditional dairy plant.

Item	total psychrotrophic count	<i>S. aureus</i> count
Incoming raw milk	$3.2 \times 10^4 \pm 1.7 \times 10^3$	$5.6 \times 10^3 \pm 6.9 \times 10^2$
Cheese vat	$4.5 \times 10^5 \pm 6.4 \times 10^4$	$2.5 \times 10^3 \pm 1.5 \times 10^3$
Cheese mould	$7.4 \times 10^4 \pm 2.7 \times 10^4$	$2.4 \times 10^3 \pm 1.7 \times 10^3$
Wooden table	$6.8 \times 10^4 \pm 2.3 \times 10^4$	$1.8 \times 10^3 \pm 3.7 \times 10^3$
Cheese handler	$5.8 \times 10^4 \pm 3.6 \times 10^4$	$9.4 \times 10^4 \pm 5.6 \times 10^3$
Water supply	$3.7 \times 10^5 \pm 1.9 \times 10^4$	$6.5 \times 10^2 \pm 1.7 \times 10^2$
Wall	$8.7 \times 10^4 \pm 3.4 \times 10^4$	$8.5 \times 10^3 \pm 2.3 \times 10^3$
Plastic package	$6.8 \times 10^4 \pm 8.5 \times 10^3$	$3.6 \times 10^3 \pm 1.6 \times 10^3$
Soft cheese	$5.9 \times 10^7 \pm 6.3 \times 10^6$	$7.8 \times 10^4 \pm 4.3 \times 10^3$

The most contaminated point with total psychrotrophic count was in cheese vat followed by water supply. The highest *S aureus* count was found in cheese handler followed by the wall.

Table 3: Total enterococci count and Total yeast and mold count during processing steps of white soft cheese in traditional dairy plant.

Item	Enterococci count	total yeast and mold count
Incoming raw milk	$4.5 \times 10^2 \pm 1.4 \times 10^2$	$8.3 \times 10^2 \pm 2.3 \times 10$
Cheese vat	$7.8 \times 10^2 \pm 2.4 \times 10^2$	$6.4 \times 10^2 \pm 1.8 \times 10^2$
Cheese mould	$6.4 \times 10^2 \pm 2.8 \times 10^2$	$5.8 \times 10^3 \pm 1.3 \times 10^2$
Wooden table	$2.8 \times 10^4 \pm 1.3 \times 10^2$	$3.4 \times 10^3 \pm 1.6 \times 10^2$
Cheese handler	$7.2 \times 10^2 \pm 2.5 \times 10^3$	$6.8 \times 10^2 \pm 2.3 \times 10^2$
Water supply	$1.3 \times 10^3 \pm 4.7 \times 10^2$	$5.9 \times 10^2 \pm 1.7 \times 10^2$
Wall	$7.3 \times 10^2 \pm 3.2 \times 10^2$	$6.6 \times 10^2 \pm 1.9 \times 10^2$
Plastic package	$4.5 \times 10^2 \pm 1.1 \times 10^2$	$8.9 \times 10^2 \pm 2.6 \times 10^2$
Soft cheese	$7.5 \times 10^4 \pm 2.4 \times 10^3$	$6.3 \times 10^4 \pm 4.2 \times 10^3$

The most contaminated point with Enterococci was wooden table followed by water supply the most contaminated point with yeast and mold was cheese mould followed by wooden table.

DISCUSSION

Most of the locally manufactured dairy products are liable to be contaminated with different types of microorganisms from different sources including steps of manufacture. For the production of dairy products of high microbiological quality, all surfaces of dairy equipment must be properly cleaned and sterilized. It is better and advisable to apply sanitation system, in the dairy plants and factories, for all surfaces on which milk comes in contact during manufacturing.

The microbial quality and safety of Egyptian white soft cheese, is the major area of concern for producers, public health authorities and consumers. It depends on the types of microorganisms introduced from raw milk, efficiency of processing and the hygienic practice applied in small or big dairy plant or informal producers. Handling of milks during cheese manufacture play an important role in the proliferation of microbial flora and consequently impair its utility and render the product unfit for human consumption, raw milk should be purchased from inspected and approved suppliers and should be stored and distributed under conditions that prevent microbial growth and contamination (EL-Baradei *et al.*, (2007).

The data in Table 1 revealed that the mean total mesophilic bacterial count during processing steps of white soft cheese were $3.2 \times 10^5 \pm 1.3 \times 10^4$, $4.6 \times 10^5 \pm 1.9 \times 10^4$, $9.4 \times 10^6 \pm 3.7 \times 10^4$, $8.5 \times 10^5 \pm 4.6 \times 10^3$ and $7.5 \times 10^5 \pm 6 \times 10^4$ for incoming raw milk, Cheese vat, Cheese mould, Wooden table and Cheese handler respectively. While the total mesophilic bacterial counts for Water supply, Wall, Plastic package and Soft cheese were $9.4 \times 10^5 \pm 1.7 \times 10^3$, $5.8 \times 10^5 \pm 4.8 \times 10^4$, $2.1 \times 10^6 \pm 3.6 \times 10^4$ and $3.9 \times 10^7 \pm 2.8 \times 10^5$, respectively. The highest total mesophilic bacterial count was found in Cheese mould by $9.4 \times 10^6 \pm 3.7 \times 10^4$ cfu/cm² followed by Plastic package with $2.1 \times 10^6 \pm 3.6 \times 10^4$ cfu/ cm². These results similar to that obtained by (Abdou, 1990). Total counts of bacteria are the most useful indicator for the microbiological status of the cheese. A high viable count often Indicates contamination of raw material, unsatisfactory sanitation, or unsuitable time and temperature during storage and/or Production. (Mossel, 1983). The presence of high total mesophilic bacterial count in raw milk indicates serious faults in production and handling or may be due to lack of cooling facilities during transportation (Mehari and Gashe, 1990).

Also, Table 1 showed that The total coliforms count during processing steps of white soft cheese were $4.2 \times 10^3 \pm 7.2 \times 10^2$, $6.1 \times 10^4 \pm 1.11 \times 10^3$, $2.1 \times 10^2 \pm 0.28 \times 10$, $6.2 \times 10^2 \pm 3.5 \times 10$ and $5.6 \times 10^3 \pm 1.8 \times 10^2$ for incoming raw milk, cheese vat, cheese mould, wooden table and cheese handler, respectively.

While, the total coliforms count for Water supply, Wall, Plastic package and Soft cheese were $3.9 \times 10^2 \pm 1.6 \times 10^2$, $5.7 \times 10^2 \pm 4.3 \times 10^2$, $3.8 \times 10^2 \pm 1.4 \times 10^2$ and $2.4 \times 10^2 \pm 1.3 \times 10$ respectively.

The most contaminated point with coliforms was cheese vat with a mean count of $6.1 \times 10^4 \pm 1.11 \times 10^3$ followed by cheese handler with mean count of $5.6 \times 10^3 \pm 1.8 \times 10^2$. In recent years attention paid toward coliform bacteria because of their public health importance, faecal coliforms are widely distributed in nature. They gain entry to milk and milk products through the water supply, equipment, unhygienic conditions of production and handling (Hafez, 1984).

All examined white soft cheese samples not complied with Egyptian standards (2005) which stated that coliform count should not be more than 10 cfu/ g.

The results in Table 2 showed that the total psychrotrophic count during processing steps of white soft cheese were $3.2 \times 10^4 \pm 1.7 \times 10^3$, $4.5 \times 10^5 \pm 6.4 \times 10^4$, $7.4 \times 10^4 \pm 2.7 \times 10^4$, $6.8 \times 10^4 \pm 2.3 \times 10^4$ and $5.8 \times 10^4 \pm 3.6 \times 10^4$ for incoming raw milk, cheese vat, cheese mould, wooden table and cheese handler, respectively. Mean while total psychrotrophic count for water supply, wall, plastic package and soft cheese were $3.7 \times 10^5 \pm 1.9 \times 10^4$, $8.7 \times 10^4 \pm 3.4 \times 10^4$, $6.8 \times 10^4 \pm 8.5 \times 10^4$ and $5.9 \times 10^7 \pm 6.3 \times 10^6$ respectively.

The most contaminated point with total psychrotrophic count was cheese vat with a mean count of $4.5 \times 10^5 \pm 6.4 \times 10^4$ followed by water supply with a mean count of $3.7 \times 10^5 \pm 1.9 \times 10^4$. Psychrotrophs in general found in water and soil and introduced in milk and milk products through these sources and established on milk contact surfaces and drain in the processing plants. Psychrotrophic bacteria are responsible for many undesirable changes in flavor and body texture of dairy products. (APHA, 1992). Psychrotrophs produce extracellular thermostable proteolytic and lipolytic enzymes which responsible for off flavor and yield loss in cheese manufacture (Prieto *et al.*, 2002).

Table 2 revealed that the *S. aureus* counts during processing steps of white soft cheese were $5.6 \times 10^3 \pm 6.9 \times 10^2$, $2.5 \times 10^3 \pm 1.5 \times 10^3$, $2.4 \times 10^3 \pm 1.7 \times 10^3$, $1.8 \times 10^3 \pm 3.7 \times 10^3$ and $9.4 \times 10^4 \pm 5.6 \times 10^3$ for incoming raw milk, cheese vat, cheese mould, wooden table and cheese handler, respectively. Also we found that the *S. aureus* counts during processing steps of soft cheese were $6.5 \times 10^2 \pm 1.7 \times 10^2$, $8.5 \times 10^3 \pm 2.3 \times 10^3$, $3.6 \times 10^3 \pm 1.6 \times 10^3$ and $7.8 \times 10^4 \pm 4.3 \times 10^3$ for water supply, wall, plastic package and soft cheese, respectively.

The highest *S aureus* count was found in cheese handler by $9.4 \times 10^4 \pm 5.6 \times 10^3$ followed by the wall with a mean count of $8.5 \times 10^3 \pm 2.3 \times 10^3$.

The presence of large number of *S. aureus* in dairy products is considered a good indicator of personal hygiene of factory workers with respiratory infections and suppurative lesions as boils (Kamat *et al.*, 1991). *S. aureus* can gain access to milk either by direct excretion from udders with clinical or subclinical staphylococcal mastitis or by contamination from the environment during handling and processing of raw milk (Peles *et al.*, 2007). *Staphylococcus aureus* may be the main cause of several food intoxication outbreaks for their production of heat stable enterotoxins (ICMSF, 1996). All examined white soft cheese samples not complied with the Egyptian standards (2005) which stated that white soft cheese should be free from any pathogenic organisms.

Table 3 showed that the Enterococci counts during processing steps of white soft cheese were $4.5 \times 10^2 \pm 1.4 \times 10^2$, $7.8 \times 10^2 \pm 2.4 \times 10^2$, $6.4 \times 10^2 \pm 2.8 \times 10^2$, $2.8 \times 10^4 \pm 1.3 \times 10^3$ and $7.2 \times 10^2 \pm 2.5 \times 10^2$ For incoming raw milk, cheese vat, cheese mould, wooden table and cheese handler, respectively. Also we found that the Enterococci counts were $1.3 \times 10^3 \pm 4.7 \times 10^2$, $7.3 \times 10^2 \pm 3.2 \times 10^2$, $4.5 \times 10^2 \pm 1.1 \times 10^2$ and $7.5 \times 10^4 \pm 2.4 \times 10^3$. For water supply, wall, plastic package and soft cheese, respectively. The most contaminated point with Enterococci was wooden table with a mean value of $2.8 \times 10^4 \pm 1.3 \times 10^2$ followed by water supply with a mean count of $1.3 \times 10^3 \pm 4.7 \times 10^2$. Enterococci are widely distributed in nature, they gain entry into milk and milk products through the water supply, equipment and unhygienic conditions of production and handling. They have been incriminated as direct or indirect cause of food born infection and food poisoning outbreaks (Garg and Mital, 1991).

The high count of Enterococci may be due to inadequate heat treatment and bad conditions of production and unsanitary handling of the product, their presence in dairy products was unaccepted as the most passive indicator of fecal contamination.

It is evident from Table 3 that the total yeast and mold count during processing steps of white soft cheese were $8.3 \times 10^2 \pm 2.3 \times 10^1$, $6.4 \times 10^2 \pm 1.8 \times 10^2$, $5.8 \times 10^3 \pm 1.3 \times 10^2$, $3.4 \times 10^3 \pm 1.6 \times 10^2$ and $6.8 \times 10^2 \pm 2.3 \times 10^2$, respectively.

While, for water supply, wall, plastic package and soft cheese were $5.9 \times 10^2 \pm 1.7 \times 10^2$, $6.6 \times 10^2 \pm 1.9 \times 10^2$, $8.9 \times 10^2 \pm 2.6 \times 10^2$ and $6.3 \times 10^4 \pm 4.2 \times 10^3$ respectively. The most contaminated point with yeast and mould was cheese mould with a mean count of $5.8 \times 10^3 \pm 1.3 \times 10^2$ followed by wooden table with a mean count of $3.4 \times 10^3 \pm 1.6 \times 10^2$. The cheese is considered as an excellent medium for yeast and molds that may induce undesirable changes such as colour defects, off-flavour and actual rots (Mislivec *et al.*, 1992).

The public health significance of yeasts and moulds has been emphasized as certain types of moulds produce mycotoxins which were implicated in human cases of food poisoning and liver cancer. All examined white soft cheese samples not complied with Egyptian standards (2005) which stated that total yeast counts must be less than 400 cfu/g and total mold counts must be less than 10 cfu/g.

The microbiological quality of the milk and the good manufacturing practices will contribute to the safety of the final product, especially in cheeses where milk is not pasteurized (FDA, 2001).

All food manufactures have a responsibility to have a full knowledge of the risks involved in their process, extending from raw materials through to consumption. The hazard analysis and critical control point (HACCP) system is described as a preventive tool for dairy manufacture (Woodhall, 1989).

RECOMMENDATIONS and CONCLUSION

Soft cheeses might represent a health risk for the consumers and considered as a possible vehicle of infection or vehicle of transmission for well-established pathogens so the cheese made in street, farmers' home or in unlicensed factories is not safe for human consumption. More and more of proper inspection must be done on street markets, supermarkets, stores, plants, unlicensed factories of dairy products. The development and application of microbiological hazard must follow established basic principles to control presence or growth of microorganisms.

To improve the safety of these product efforts to raise awareness of the importance of hygiene barriers and raw milk quality as well as improved process control can be suggested, we can recommend that the receiving of raw milk should be carefully monitored and only obtained from suppliers apply good manufacturing practices. Also strict hygienic measures of cleaning and sanitization of all food contact surfaces and hygienic training of plant workers should be applied to avoid contamination, water supply must be clean and comply with the standard requirements, prevention of environmental contamination, good cleaning and sanitizing of food processing is essential to produce safe and high quality cheese. Good conditions of hygiene should be maintained throughout cheese manufacture until consumption to prevent contamination. HACCP – based risk assessment, good manufacturing practice and ISO 22000 food safety should be implemented for all stages of manufacture in order to produce safe and good quality dairy product.

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تقييم المخاطر الميكروبية أثناء إنتاج الجبن الأبيض في احد مصانع الألبان التقليدية

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الجبن الأبيض المصنع محليا من منتجات الألبان واسعة الانتشار لما يحويه من الكثير من العناصر الغذائية اللازمة لاحتياجات جسم الإنسان. وعلى الرغم من ذلك فإن خطورة منتجات الألبان غالبا تكون خطورة ميكروبيولوجية لذلك فهي الأداة في تقييم قبول المنتج. ومنتجات الألبان تكون أوساط غذائية عالية الجودة بالنسبة للميكروبات وبالتالي يؤدي ذلك إلى فساد المنتج. والغرض من هذا العمل هو تحديد نقاط التحكم الحرجة أثناء تصنيع الجبن الأبيض ومحاولة تقليل هذه الخطورة وتم اخذ عينات من مراحل التصنيع من كل من اللبن الخام وحوض التصنيع والتعليق والمنضدة الخشبية وايدى العمال ومصدر المياه والجدران وبلاستيك التغليف والمنتج النهائي (الجبن الأبيض) وقد تم إجراء الاختبارات الميكروبيولوجية الآتية: العد البكتيري الكلى - عد المجموعة القولونية - العد الكلى للميكروبات المحبة للبرودة - العد الكلى للميكروبات المكور العنقودي الذهبي - عد الميكروبات السحبية المعوية - العد الكلى للفطريات والخمائر وأظهرت نتائج الاختبارات الميكروبيولوجية ما يلي:- العد البكتيري الكلى كانت أعلى نقاط التلوث أثناء تصنيع الجبن الأبيض هي التعليق ويليها بلاستيك التغليف وبالنسبة لعد المجموعة القولونية كانت أعلى نقاط التلوث أثناء تصنيع الجبن الأبيض هي حوض التصنيع ويليها ايدى العمال وكانت أعلى نقاط التلوث بالميكروبات المحبة للبرودة أثناء تصنيع الجبن الأبيض هي حوض التصنيع ويليها مصدر المياه وبالنسبة للعد الكلى للميكروبات المكور العنقودي الذهبي كانت أعلى نقاط التلوث أثناء تصنيع الجبن الأبيض هي ايدى العمال ويليها الجدران وكان عد الميكروبات السحبية المعوية أعلى في المنضدة الخشبية ويليها مصدر المياه. وكانت أعلى نقاط التلوث بالفطريات والخمائر أثناء تصنيع الجبن الأبيض هي التعليق ويليها المنضدة الخشبية. لذلك توصى الدراسة بالتعرف على النقاط الحرجة في مراحل تصنيع الجبن الأبيض وإتباع الطرق الصحية للتحكم فيها في المصانع المختلفة عن طريق تطبيق نظام الهاسب والغذاء الآمن أثناء مراحل التصنيع والتخزين والتسويق للحصول على منتج آمن وذو جودة عالية.