

## EXPERIMENTAL STUDIES ON COCCIDIA (*EIMERIA TENELLA*) INFECTED BROILER TREATED WITH AQUEOUS EXTRACT OF OYSTER MUSHROOM

GHADA A.A.\*; DOAA A.H.\*\*; REHAB R.A.\*\*\* and RANIA I.M.\*\*\*\*

\* Department of Poultry Diseases, Animal Health Research Institute, Agriculture Research Center, Dokki, Giza, Egypt, Al-Mansoura Provincial Lab.

\*\* Department of Mycology, Animal Health Research Institute, Agriculture Research Center, Dokki, Giza, Egypt, Al-Mansoura Provincial Lab.

\*\*\* Department of Parasitology, Animal Health Research Institute, Agriculture Research Center, Dokki, Giza, Egypt, Al-Mansoura Provincial Lab.

\*\*\*\* Department of Pathology, Animal Health Research Institute, Agriculture Research Center, Dokki, Giza, Egypt, Al-Mansoura Provincial Lab.

Email: [dr.ahmedsob7y@gmail.com](mailto:dr.ahmedsob7y@gmail.com)

---

### ABSTRACT

---

**Received at: 24/8/2014**

**Accepted: 19/10/2014**

The present work was undertaken to determine aqueous extract of mushroom (*Pleurotus ostreatus*) as an alternative to other coccidiosis control measures would result in improving body weight, oocyst reduction and improving pathological lesions of cecum. A total of 140 broiler chicks of 14 days age were divided into 7 groups (each contain 20 chicks). The 1<sup>st</sup> group was not infected with *Eimeria tenella* and didn't take any medicine or additives and kept as control, while the 2<sup>nd</sup> group was infected with *Eimeria tenella* at a rate of 50,000 sporulated oocyst/ml / bird at 14<sup>th</sup> day of age. Third group was infected with *Eimeria tenella* as mentioned before at 14<sup>th</sup> day of age and treated with aqueous extract of mushroom (200 mg/ml) at day 6 post infection and continued for 7 days consecutively. Fourth group was infected with *E. tenella* as mentioned before and treated with amprolium (200 mg/ml) at 6 day post infection and continued for 7 days consecutively. Fifth group was infected with *E. tenella* as mentioned before and treated with both aqueous extract of mushroom (200 mg/ml) and amprolium (200 mg/ml) at 6 day post infection, for 7 days. Sixth group was non infected with *E. tenella* but received aqueous extract of mushroom at 14<sup>th</sup> day of age (200 mg/ml) and continued till the end of experiment (at 35 days of age). Seventh group not infected with *E. tenella* but received amprolium (200 mg/ml) at 14<sup>th</sup> day of age and continued till the end of experiment (the period of experiment is three weeks). Body weight was recorded weekly, oocysts counted at 21, 28 and 35 days of age, and pathological lesions were studied at 28 and 35 days of age. The obtained data revealed that chicks of the 2<sup>nd</sup> group showed a significant decrease in body weight, while third, fifth and sixth groups showed a significant increase in body weight which appeared clearly at 35day of age. Fifth group posses the highest body weight followed by chicks of sixth group then third group. The highest oocyst count was recorded in chicks of the 2<sup>nd</sup> group while the lowest oocyst count was recorded in 5<sup>th</sup>, 4<sup>th</sup> and 3<sup>rd</sup> groups. Regarding pathological lesions, the infected and untreated 2<sup>nd</sup> group showed severe necrosis of ceca with destruction and desquamation of the lining epithelium. The necrotic mucosa was heavily infiltrated or replaced with lymphocytes, macrophages and few heterophiles. The pathological lesions were less in the 3<sup>rd</sup> group (infected and treated with mushroom) and nearly absent in both 4<sup>th</sup> group (infected and treated with amprolium) and 5<sup>th</sup> group (infected and treated with both mushroom and amprolium).

---

**Key words:** *Eimeria Tenella*, Broiler, Mushroom extract.

---

### INTRODUCTION

Coccidiosis is an acute to chronic infectious disease caused by protozoal parasites of genus *Eimeria* which multiply in intestinal mucosa of

chickens and produce severe tissue damage resulting in bloody diarrhea, reduce growth, weight and increase susceptibility to other pathogens. So, avain coccidiosis is a fatal disease which cost the industry millions of dollars annually (Champman and Shirley

2003, Lilleh *et al.*, 2004). The cost associated with prevention via drugs in the feed, adds to the economic hardship of producer. Moreover, the increased drug resistant problems occurring in poultry production has heightened public concern. In some countries the use of drug in production has been severely regulated or banned altogether there for alternative control methods are being researched with great intensity, one such alternative receiving much interest is mushroom. Mushroom a natural health promoting fungi such as (*Pleurotus ostreatus*) and (*Ganoderma lucidum*) used as food supplements and medicines to improve various parameters of human and animal health and immune function in certain disease conditions (Chang and Mshigeni, 2001, Anthony and Joyce, 2007 and Fasuyi, 2007).

Mushroom like probiotics are natural ingredients that contain bioactive chemical substances or polysaccharides, protein, crude fibers, unsaturated fat, minerals as (potassium, phosphorus, calcium, iron and zinc), vitamins, essential amino acids and organic acids that can be used as a good source of supplements and medicine to promote health and production (Jang and Brimingham, 1992; Chang and Mshigeni, 2001; Ogbe *et al.*, 2005; Anonymous, 2007; Ezeokeke, 2008 and Selegean *et al.*, 2009). Mushroom stabilise microflora in GIT and prevents colonization of host cells by pathogens, also stimulates non specific host immune response or phagocytosis by macrophages (Sundu *et al.*, 2006). There has been a recent upsurge of interest in mushroom not only as a healthy food which is rich in protein (Baross *et al.*, 2008), but also as a source of biologically active compounds of medicinal value which include complementary medicinal dietary supplements for anticancer (Cheung *et al.*, 2003), antioxidant (Chang and Miles, 2004), antimicrobial (Lindequist *et al.*, 1990), antiviral (Brandt and Piraino, 2000 and Mothana *et al.*, 2003) and anti inflammatory (Kim *et al.*, 2003 and 2004).

In addition, it has immunopotentiating (Reshetinkov *et al.*, 2001) hypocholesterolemic, (Ishikawa *et al.*, 1984) and antioocidal effect against *Eimeria tenella* (Ogbe *et al.*, 2009; Naphade *et al.*, 2010; Willis *et al.*, 2012 and Hossain *et al.*, 2013).

Infection with *Eimeria* is known to stimulate a protective immune response in chicken (Yun *et al.*, 2000). The polysaccharide extract from mushroom (*Pleurotus ostreatus*) was shown to have immunomodulating effects in chicken (Selegean *et al.*, 2009). Mushroom can reduce *Eimeria tenella* oocysts output in infected broilers, improve body weight gain and hematological changes that may occur (Ogbe *et al.*, 2009).

This study was carried out to investigate the performance and health promoting effects of aqueous

extracts of mushroom (*Pleurotus ostreatus*) on broiler chicks infected with *Eimeria tenella*.

## **MATERIALS and METHODS**

**1- Experimental birds:** One hundred and forty chicks of 14 day age were obtained from a commercial hatchery. The chicks were weighed, divided into 7 groups (20 chicks each) and housed on wire cages labeled 1 to 7. All groups of birds were fed with standard ration.

**2- Preparation of aqueous extract of *Pleurotus ostreatus*** (according to Ogbe *et al.*, 2009): Mushroom was washed in distilled water, sun dried then ground to powder using a mortar pestle and then blended. The mushroom powder was again sun dried for about 3 hours and then stored in plastic polythene bags and kept at room temperature until required. A 20% w/v solution of aqueous extract of mushroom was prepared by soaking in hot water boiled to 100°C for 3 hours brining the concentration to 200mg/ml. The solution was sieved, solid matter discarded and the filtrate allowed to cool to room temperature before use.

### **3- Experimental infection and treatment:**

**1<sup>st</sup> group:** Control group, non infected and non treated.

**2<sup>nd</sup> group:** Infected with *Eimeria tenella* at a rate of 50,000 sporulated oocyst/ml per bird using an insulin syringe introduced directly into crop of each bird at 14<sup>th</sup> day of age. (Oocyst per gram (OPG) of feces was counted following McMaster technique.

**3<sup>rd</sup> group:** Infected with *Eimeria tenella* as mentioned before at 14<sup>th</sup> day of age and treated with aqueous extract of mushroom (200mg/ml) by day 6 post infection and continued for 7 days consecutively.

**4<sup>th</sup> group:** Infected with *Eimeria tenella* at 14<sup>th</sup> day of age as mentioned before and treated with amprolium (200mg/ml drinking water) by 6 day post infection and continued for 7 days consecutively.

**5<sup>th</sup> group:** Infected with *Eimeria tenella* as mentioned before at 14<sup>th</sup> day of age, by 6 day post infection and treated with both aqueous extract of mushroom (200mg/ml) and amprolium (200mg/ml) given for 7 days consecutively.

**6<sup>th</sup> group:** Non infected with *Eimeria tenella* but received aqueous extract of mushroom (200mg/ml) at 14<sup>th</sup> day of age and continued till the end of experiment.

**7<sup>th</sup> group:** Non infected with *Eimeria tenella* but received amprolium (200mg/ml) at 14<sup>th</sup> day of age and continued till the end of experiment.

**1 - Determination of body weight:** Body weight of broilers in all groups was monitored weekly using a weighting balance every morning prior to feeding.

**2 - Collection of fecal sample and laboratory examination:** The faeces of broiler chicks were

collected at 21,28,35 day of age in plastic bags for parasitological examination. Oocysts per gram (OPG) were counted by McMaster's slide (Hodgson, 1970).

**3 - The lesion score of *E.tenella* infestation:** was carried out according to (Johanson and Reid 1970).

**4 - Pathological examination:** Clinical signs and post mortem findings were recorded on the experimental birds before and after the day of sacrifice (1 and 2 weeks post infection or post treatment). Specimens from the ceci were collected and fixed in 10% buffered neutral formalin solution, dehydrated in gradual ethanol (70-100%), cleared in xylene and embedded in paraffin, five micron sections were prepared and then routinely stained with hematoxylin and eosin (H&E) (Bancroft and Gambl 2008) and then examined microscopically.

**5 - Statistical analysis:** The obtained data were analyzed using the liner model programs of SAS (1990). The difference among means were tested using Duncan Multiple range test (Duncan, 1955).

## RESULT

**1. Body weight record:** The recurrent results in table (1) showed that at 21 day of age (before treatment), second group (infected, untreated chicks) showed a significant decrease in body weight ( $465.50 \pm 1.5$ ) compared to control 1<sup>st</sup>group (uninfected, untreated)

( $540.50 \pm 4.5$ ) and compared to all another groups (3<sup>rd</sup>,4<sup>th</sup>,5<sup>th</sup>,6<sup>th</sup>,7<sup>th</sup>) which showed non significant difference between each other and between control 1<sup>st</sup> group. At 28 days of age (one week PT.), 2<sup>nd</sup> group (infected, untreated) showed a significant decrease in body weight ( $590.60 \pm 9.6$ ) in comparison with control 1<sup>st</sup>group ( $879.90 \pm 13.6$ ) and in comparison with all another groups. Both 5<sup>th</sup> group (infected, treated with mushroom and amprolium) and 6<sup>th</sup> group (uninfected, treated with mushroom) recorded the highest body weight ( $899.60 \pm 16.9, 899.60 \pm 8.5$ ), they showed non significant difference between each other and between control 1<sup>st</sup>group ( $879.90 \pm 13.6$ ), followed by 3<sup>rd</sup> group (infected, treated with mushroom) ( $886.20 \pm 3.9$ ) then 7<sup>th</sup>group (uninfected, treated with amprolium) ( $870.40 \pm 3.9$ ) which showed non significant difference between each other and between control 1<sup>st</sup> group. At 35days of age (2week PT.), 2<sup>nd</sup> group showed significant decrease in body weight ( $800.70 \pm 8.5$ ) as compared to control 1<sup>st</sup>group ( $1189.5 \pm 14.4$ ) and in comparison with all another groups. Fifth Group recorded the highest body weight ( $1378.5 \pm 18.5$ ) which showed significant increase as compared to control 1<sup>st</sup>group and all another groups, followed by 6<sup>th</sup> group ( $1303.4 \pm 8.1$ ) then 3<sup>rd</sup> group ( $1299.7 \pm 2.4$ ) which showed significant increase in body weight in comparison with control 1<sup>st</sup>group. Whereas 7<sup>th</sup> and 4<sup>th</sup> groups ( $1220.4 \pm 5.2, 1208.8 \pm 6.9$ ) showed non significant difference in body weight between each other and between control 1<sup>st</sup> group.

**Table 1:** Mean values of body weight of the 7 groups of broilers under experiment.

Parameters	1 <sup>st</sup> group (unchallenged ,untreated )	2 <sup>nd</sup> group (challenged ,untreated)	3 <sup>rd</sup> group (challenged, treated with mushroom)	4 <sup>th</sup> group (challenged, treated with amprolium)	5 <sup>th</sup> group (challenged, treated with mushroom and amprolium)	6 <sup>th</sup> group (unchallenged , treated with mushroom)	7 <sup>th</sup> group (unchallenged ,treated with amprolium)
<b>14 days of age</b>	$321.80 \pm 14.6^d$	$360.20 \pm 2.9^b$	$363.70 \pm 2.1^{ab}$	$370.70 \pm 3.8^a$	$333.80 \pm 3.81^c$	$363.60 \pm 3.02^{ab}$	$360.30 \pm 3.8^b$
<b>21days of age</b>	$540.50 \pm 4.5^b$	$465.50 \pm 1.5^c$	$554.70 \pm 2.7^{ab}$	$558.50 \pm 3.5^{ab}$	$568.20 \pm 13.9^{ab}$	$560.10 \pm 7.6^{ab}$	$548.20 \pm 4.1^{ab}$
<b>28days of age</b>	$879.90 \pm 13.6^{abc}$	$590.60 \pm 9.6^d$	$886.20 \pm 3.9^{ab}$	$864.20 \pm 5.9^{bc}$	$899.60 \pm 16.9^a$	$899.60 \pm 8.5^a$	$870.40 \pm 3.9^{ab}$
<b>35days of age)</b>	$1189.5 \pm 14.4^c$	$800.70 \pm 8.5^d$	$1299.7 \pm 2.4^b$	$1208.8 \pm 6.9^c$	$1378.5 \pm 18.5^a$	$1303.4 \pm 8.1^b$	$1220.4 \pm 5.2^c$

Value are means  $\pm$  standard error, means with different letters at the raw differ significantly at (P <0.05).

**1 - Oocysts per gram count:** The analysis of results in table (2) showed that, at 21 day of age (befor treatment): The oocysts output was (47100±458.25) 2<sup>nd</sup> group, (45000±1414.21) 3<sup>rd</sup> group, (45700±422.95) 4<sup>th</sup> group and

2 - (44200±840.64) 5<sup>th</sup> group. At 28 day of age (one week post treatment), 2<sup>nd</sup> group showed high significant increase in OPG counts (70000±1626.17) while 3<sup>rd</sup>, 4<sup>th</sup>, 5<sup>th</sup> groups showed significant decrease in oocysts count (7553±2.33, 2567±1.86, 2067±1.7)

respectively as compared to control untreated 2<sup>nd</sup> group, at the same time 3<sup>rd</sup>, 4<sup>th</sup> and 5<sup>th</sup> groups showed non significant difference in oocyst count between each other. At 35 days of age (2week post treatment), 2<sup>nd</sup> group showed significant increase in oocysts count (60000±4216.37) while all treated groups (3<sup>rd</sup>, 4<sup>th</sup>, 5<sup>th</sup>) showed significant decrease (50±2.58, 2±0.36, 1±0.2) respectively as compared to control 2<sup>nd</sup> group, as well as, these groups showed non significant difference in oocyst count between each other.

**Table 2:** Mean values of OPG count of broilers

Parameters	1 <sup>st</sup> group (unchallenged, untreated)	2 <sup>nd</sup> group (challenged, untreated)	3 <sup>rd</sup> group (challenged, treated with mushroom)	4 <sup>th</sup> group (challenged, treated with amprolium)	5 <sup>th</sup> group (challenged, treated with mushroom and amprolium)	6 <sup>th</sup> group (unchallenged, treated with mushroom)	7 <sup>th</sup> group (unchallenged, treated with amprolium)
<b>21 days of age</b>	0	47100± 458.25 <sup>c</sup>	45000± 1414.21 <sup>a</sup>	45700± 422.95 <sup>a</sup>	44200± 840.64 <sup>a</sup>	0	0
<b>28 days of age</b>	0	70000± 1626.17 <sup>a</sup>	7553± 2.33 <sup>b</sup>	2567± 1.86 <sup>b</sup>	2067± 1.7 <sup>b</sup>	0	0
<b>35 days of age</b>	0	60000± 4216.37 <sup>b</sup>	50± 2.58 <sup>c</sup>	2± 0.36 <sup>c</sup>	1± 0.2 <sup>c</sup>	0	0

Value are means ± standard error, means with different letters at the raw differ significantly at (P <0.05).

**3. Mortality rate and morbidity:**

No mortality occurred in any of the experimental groups till the end of the experiment while on day 5 PI. all the birds in the infected groups (2, 3, 4 and 5) appeared dull, weak, loss of appetite, their feces became bloody and watery and *Eimeria tenella* oocysts were detected in their feces. These symptoms nearly disappeared in chicks of third group at (one week post treatment) and disappeared completely at (two week post treatment.). While in chicks of fourth and fifth groups the symptoms disappeared completely at (one week post treatment). On the other hand, in chicks of second group the symptoms continued till (two week PI.) then decreased gradually.

**4. Pathological findings**

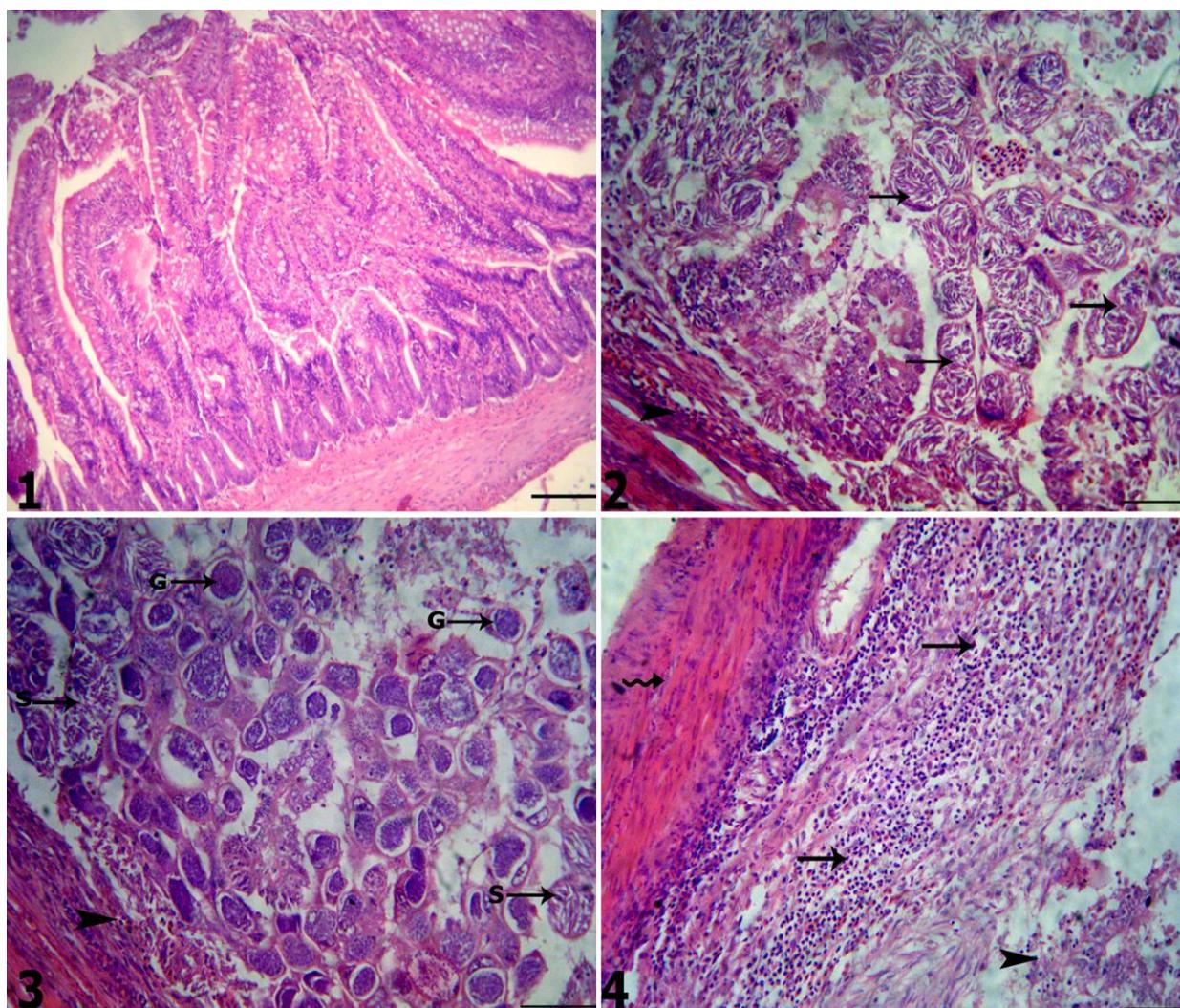
**The ceca of control (group 1):** “Non-infected, non-treated” showed neither gross nor microscopic abnormalities along the experimental periods (Fig 1). However, the ceca of (group 2): “Infected, non-treated” macroscopically showed extensive hemorrhages in the cecal lumen and petechiation on the mucosa and serosa, particularly at 1 week PI and hemorrhagic cecal core on 2 weeks PI. Microscopically, the cecum revealed severe necrosis, destruction and desquamation of the lining epithelium with the presence of developmental stages of *Eimeria* including mature schizonts (Fig 2) and gamonts (Fig

3) in the enterocytes and lumens with extensive extravasated erythrocytes, 1 and 2 weeks PI. Sometimes, the necrotic mucosa was heavily infiltrated or replaced with lymphocytes and few heterophils besides hyalinization of the muscular coat (Fig 4). Hemorrhages, edema and necrosis were seen in the submucosa particularly near the muscular layer. Almost all the epithelial lining of the cecal mucosa were invaded with different developmental stages of coccidia (schizonts and gamonts), 2 weeks PI (Fig 5). The submucosa was edematous and severely infiltrated with lymphocytes, macrophages and few heterophils.

**The ceca of group (3):** “Infected and treated with Mushroom” showed slight lowering in the lesions of group (2). Bloody contents and petechial hemorrhages on the mucosa were also visualized. Microscopically, the developmental stages of *Eimeria* were evident in the lining epithelium of the cecum similar to those described in the infected chickens besides necrotic mucosa and lymphocytes infiltrations, 1 and 2 weeks PT (Fig 6). The mucosa of the cecum was slightly intact and the submucosa was infiltrated with lymphocytes and few macrophages, 2 weeks PT. Some hyperplastic changes were seen in the cecal epithelium and crypts. These findings in this group were presented as regenerative attempts. The lamina propria and

submucosa showed edema and focal fibrosis. Mild hyperplasia of the lymphoid follicles of cecal tonsils was also seen. Meanwhile **the ceca of group (4):** “Infected and treated with amprolium” showed nonspecific changes, particularly at 2 weeks PT. Microscopically, the ceca showed intact mucous membrane with few developmental stages of Eimeria with numerous lymphocytes and macrophages infiltrations, particularly in the crypts, 1 week PT (Figs 7 and 8). Partial desquamation of the lining epithelium was also detected. However, the ceca at 2 weeks PT showed intact intestine with mild mucinous degeneration and few round cells infiltrations. Severe hyperplasia in the lining epithelium was noticed with no evidence of coccidial stages (Fig 9) besides congested blood vessels and thick eosinophilic membrane coated the luminal surface of the lining epithelium. **The ceca of group (5):** “Infected and

treated with both amprolium and Mushroom” were apparently normal particularly at 2 weeks PT. Microscopically, the lesions of such group were completely ameliorated with intact mucosa, complete absence of the developmental stages of Eimeria and few lymphocytes infiltrations in the submucosa (Fig 10). Few birds, sacrificed 1 week PT, showed few) degenerated or necrotic gamonts in the lining epithelium of crypts (Fig 11). The cecal mucosa was focally regenerated with hyperplastic lining epithelium in both sacrificed periods. The submucosa and lamina propria were infiltrated with round cells of mostly lymphocytes besides hyperplasia in the lymphocytes of the cecal lymphoid follicles and tonsils (Fig 12). The ceca of groups (6,7): Showed neither gross nor microscopic abnormalities along the experimental periods.



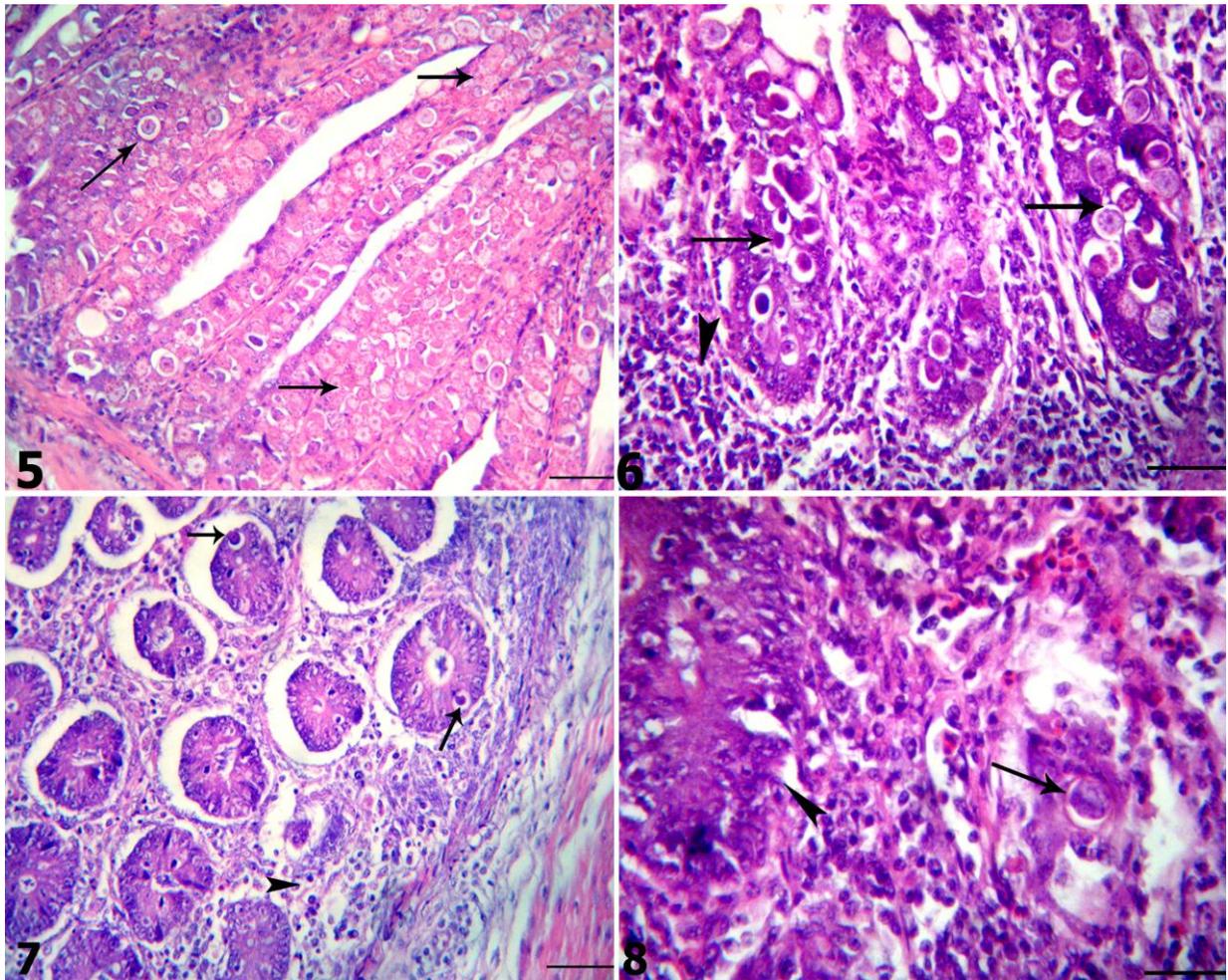
**Figs. (1-4):** Cecum from groups:

**Fig.(1), (group1):** shows normal intact mucosa and lymphoid follicles. H&E x 100.

**Fig.(2), (group 2):** shows several schizonts (arrows) in the enterocytes. H&E x 400.

**Fig.(3), (group 2):** shows numerous gamonts (arrows) in the enterocytes. H&E x 400.

**Fig.(4), (group2):** shows severe necrosis and lymphocytic infiltrations (arrow) with hyalinization of muscular coat (irregular arrow). H&E x 200.



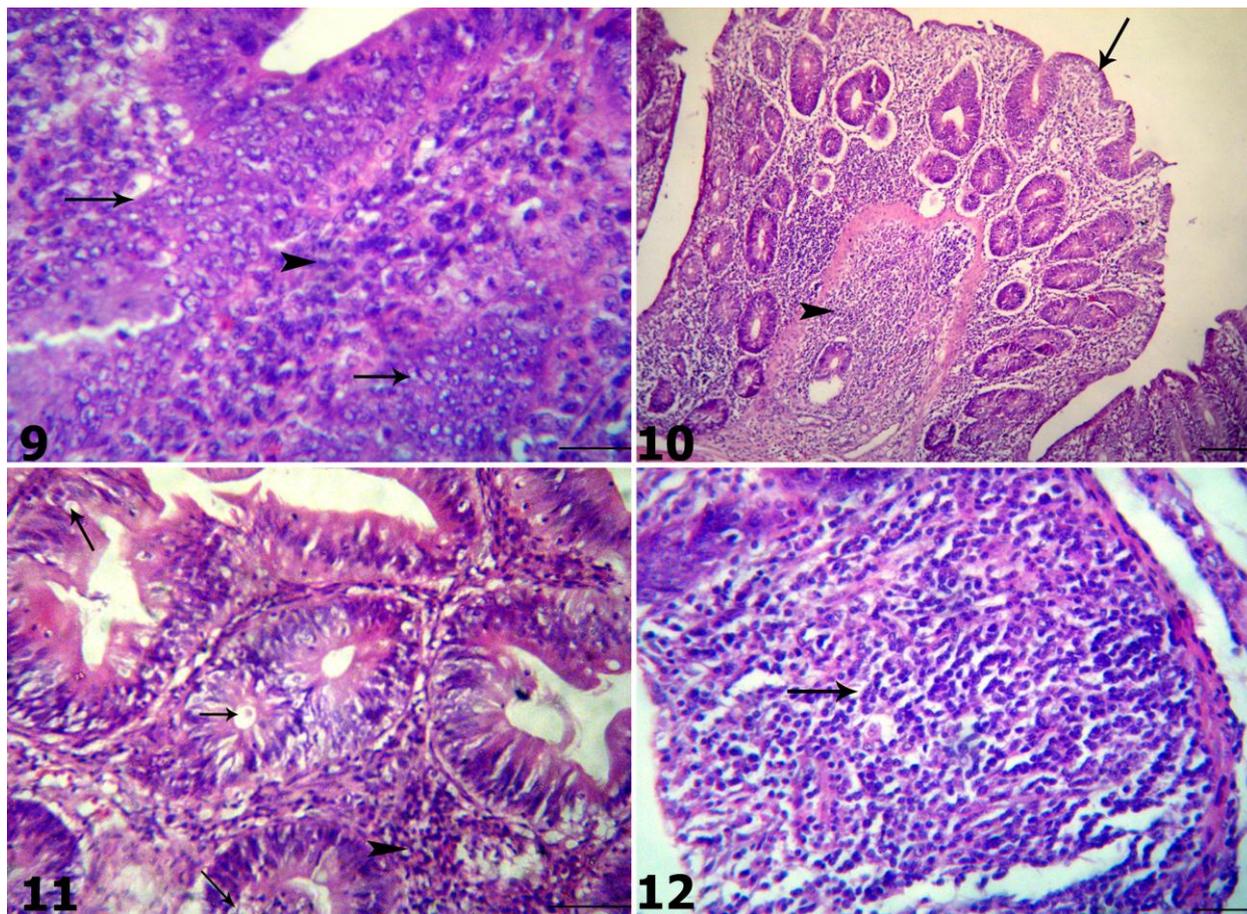
**Figs. (5-8):** Cecum from groups:

**Fig.(5), (group 2):** shows developmental stages in almost all the intestinal epithelium (arrows). H&E x 200.

**Fig.(6), (group 3):** shows developmental stages of *Eimeria* in the lining epithelium and numerous lymphocytes infiltrations. H&E x 200.

**Fig.(7), (group 4):** shows few developmental stages of *Eimeria* in the lining epithelium of crypts (arrows) with numerous lymphocytes and macrophages infiltrations (arrowhead). H&E x 200.

**Fig.(8):** A higher magnification of fig (7). H&E x 400.



**Figs. (9-12):** Cecum from groups:

**Fig.(9), (group 4):** shows severe hyperplasia in the lining epithelium (arrows) with no evidence of Eimeria. H&E x 400.

**Fig.(10), (group 5):** shows intact mucosa with no evidence of developmental stages (arrow) and lymphocytes infiltrations (arrowhead). H&E x 100.

**Fig.(11), (group 5):** shows few degenerated gamonts in the lining epithelium of crypts (arrows). H&E x 400.

**Fig.(12), (group 5):** shows hyperplasia in lymphocytes of the lymphoid follicles (arrow). H&E x 200.

**Scoring of histopathological findings in the ceca.**

Groups	Necrosis	Viable stages	Degenerated stages	Cecal tonsils and lymphoid follicles	Hemorrhages and congestion	Leukocytes infiltrates
1	-	-	-	N	-	-
2	++++	++++	+	Slightly edematous	++++	+++
3	+++	+++	++	Mild hyperplasia	++	++++
4	+	+	+++	N	+	+++
5	±	±	++++	Hyperplasia	±	++++
6	-	-	-	N	-	-
7	-	-	-	N	-	-

++++ Serious    +++ Severe    ++ Moderate    + Mild    ± rare    - Nil

## **DISCUSSION**

The present study investigated the anticoccidial efficacy of Oyster mushroom in relation to body weight, OPG count of faeces, morbidity, mortality, score lesion and pathological lesions. All broilers were infected (100%) by 7days PI., showed clinical signs of weakness, reduced appetite and bloody diarrhea. Regarding to the body weight, there was a significant decrease in the body weight of infected, untreated birds (2<sup>nd</sup> group); while the infected chickens which were treated with mushroom (3<sup>rd</sup> group) or with both mushroom and amprolium (5<sup>th</sup> group) showed a marked increase in body weight. Those results agreed with that obtained by (Guo *et al.*, 2003; Ogbe *et al.*, 2009 and willis *et al.*, 2012) who found that mushroom have polysaccharides that stimulating the activities of T and B lymphocytes, macrophage and natural killer cells (NK) inducing production and secretion of cytokines and complement, so it controls certain parasitic diseases. Among the reason why the treated broilers gained weight more than the non treated birds could be due to that aqueous extract of mushroom like amprolium may affect or prevent development of *E.tenella* stages. It appeared that this wild mushroom may contain compounds that are active against *E.tenella*. It was found to be non toxic in animal toxicity studies even when used at high therapeutic dose (Harkonen, 1998 and James, 2002). As well as the mushroom extract and amprolium stimulate appetite so the uninfected broilers which was treated with mushroom (6<sup>th</sup> group) or treated with amprolium (7<sup>th</sup> group) performed even high body weight.

Large numbers of oocysts were detected in the faeces of challenged birds that had received the primary infection challenge but not treated (2<sup>nd</sup> group). That data clearly showed that birds were adversely affected by this protozoan while broilers in treated groups (3, 4 and 5) showed the lowest oocyst count. Those results demonstrate the effectiveness of the mushroom in reducing the shedding of oocysts and agreed with those obtained by (Ogbe *et al.*, 2009; Willis *et al.*, 2010 and willis *et al.*, 2012) who showed that diet supplementation with aqueous extract of mushroom (FMG) exhibited a reduction in oocyst excretion and mortality in *Eimeria* challenged broiler chicks. Mushroom have bioactive compounds or polysaccharides are known to play vital roles in enhancing health. They block colonization of the intestine by pathogens, thereby improving their elimination from the body (Guo *et al.*, 2003 and Elmusharafa *et al.*, 2006) and also it contains organic acids, resin and glycosides which include steroid are known to have therapeutic use against microbes and parasites (Anon 2007; Deihk *et al.*, 2007).

Regarding to histopathological findings the cecum of infected, untreated chicks (2<sup>nd</sup> group) showed severe

necrosis, destruction and desquamation of the lining epithelium with presence development stages of *Eimeria* sometimes the necrotic mucosa was heavily infiltrated with lymphocytes, haemorrhages, oedema and necrosis were seen near the muscular layer. Those result agreed with that obtained by Soomro *et al.* (2001); Ogbe *et al.* (2005) and Enas (2011). There were reduction in those lesions in broilers infected and treated with mushroom (3<sup>rd</sup> group) and the lesions were ameliorated in broilers of (4<sup>th</sup> group) (treated with amprolium) and (5<sup>th</sup> group) (treated with both amprolium and mushroom). The regeneration of cecal mucosa was observed and there were decrease in the number of oocysts of *E. tenella* in epithelial at 2 week PT. From those results, it could be demonstrated that both amprolium and mushroom lead to improve the pathological finding induced by *E. tenella* as mushroom lead to improve innate immune responses against coccidiosis. Those finding, however, correspond to those recorded by (Naphade *et al.*, 2010 and Hossain *et al.*, 2013).

## **CONCLUSION**

Treatment with aqueous extract of oyster mushroom lead to improve body weight more than amporilum, while treatment with amprolium lead to the reduction of faecal oocysts and improve pathological lesions more than oyster mushroom. On the other hand best results were obtained when we used oyster mushroom mixed with amporilum.

## **ACKNOWLEDGMENT**

It is a great pleasure to record our independence to Prof. Dr. Mohamed Hamed Mohamed, Professor of Pathology, Faculty of Veterinary Medicine, Zagazig University for his great help throughout the period of study reading and comments on histopathological specimen.

## **REFERENCES**

- Anon, A. (2007): Mushroom may be active against fowl parasite. Thisday, 11(4280) 36.
- Anonymous, M. (2007): Mushroom may be active against fowl parasite. Thisday, 11 (4280): 9–36.
- Anthony mm, J. and Joyce, C. (2007): Proximate and nutrient composition of three types of indigenous edible wild mushroom grown in Tanzani and their utilization prospects (AJFAND). Afr. J.Food Agric. Nutr. Dev. 7(6): 1-16.
- Bancroft, J.D. and Gambl, M. (2008): Theory and practice of histological techniques 5<sup>th</sup> ed., Churchill Livingstone. New York, Londone, Philadelphia.

- Baross, C.R.; Crus, T.; Baptista, p. and Ferreira, I.C. (2008): Wild and commercial mushroom as a source of nutrients and nutraceuticals. Food Chem. Toxicol. 46, 2742.
- Brandt, C.R. and Piraino, F. (2000): Edible ectomycorrhizal Mushrooms Current knowledge and future prospects. Recent Res. Dev. Antimicrob Agents Chemother. 4, 11.
- Champman, H.D. and Shirley, M.W. (2003): The Houghton strain of *Eimeria tenella*: a review of the type strain selected for genome sequencing. Avian Pathology 31: 115-127.
- Chang, S.T. and Mshigeni, K.E. (2001): Mushroom and their human health. Their growing significance as potent dietary supplements. The university of Namibia Windhoek, pp. 1-79.
- Chang, S.T. and Miles, P.G. (2004): Mushrooms: Cultivation, Nutritional Value, Medicinal Effect, and environmental impact: FL: CRC press, (2004): PP. 27-38.
- Cheung, L.M.; Cheung, P.C.K. and Ooi, V.E. (2003): Antioxidant activity and total phenolics of edible mushroom extract. Food Chem. 81, 249.
- Deihk, K.; Ros, SP. and Mackenzie, AM. (2007): Sheanut (*vitellaria paradoxa*) meal as a feed ingredient for poultry. World's Poultry Sci. J. 63: 611-624.
- Duncan, D.B. (1955): Multiple range and multiple F-test. Biometric 11:1-42.
- Elmusharafa, M.A.; Bautista, V.; Nollet, L. and Beynen, AC. (2006): Effect of a mannanoligosaccharide preparation on *Eimeria tenella* infection in broiler chickens. International J. Poultry Sci. 5: 583-588.
- Enas, A.A. (2011): Evaluation of some anticoccidial vaccines in prevention of chickn coccidiosis Thesis of M.D. Avian and Rabbit disase Dep, Fac. of Med. Zagazig University.
- Ezeokeke, C.T. (2008): Effect of prebiotics and probiotic as growth promoters in broiler chicks Niger. J. Anim. Prod. 35(1): 162-169.
- Fasuyi, A.O. (2007): Amaranthus cruentus leaf meal as a protein supplement in broiler finisher diets hematological responses, carcass characteristics and relative organ weights (AJFAND). Afr. J. Food Agric Nutr. Dev. 7(6): 1-14.
- Guo, F.C.; Savelkoul, H.FH.; Kwakkel, R.P. and Williams, B.A. (2003): Immunoactive medicinal properties of mushroom and herb polysaccharides and their potential use in chicken diet. World's Poultry Scinces Journal 59, 427-440.
- Harkonen, M. (1998): Use of mushroom by Finns and Karelians. Intrnational of Circumpolar Health. 57: 40-55.
- Hodgson, J.N. (1970): Coccidiosis: Oocyst counting technique for coccidiostat evaluation. Exp., Parasitol. 28: 99-100.
- Hossain, M.Z.; Khan, M A.H.; Ashraf, M.M. and Islam, A.K. (2013): Role of oyster mushroom (*Pleurotus ostreatus*) against cecal coccidiosis in cobb 500 broiler chicken. J. Sci., Res. 5(1) 185-193.
- Ishikawa, Y.; Marimoto, K. and Hamasaki, T. (1984): Flavoghaucin a metabolite of Eurotium Chevalieri, its Antioxidation and Synergism with Tocopherol. J. Am. Oil chem. Soc. 61, 1864.
- James, M. (2002): Reisk Mushroom extract and Immune Support. Dynamic Practice 20: 1-5.
- Jang, SC. and Briminghan, J.M. (1992): Medicinal benefits of mushroom, Gandodroma, APPL. Microbiol. 37: 101-134.
- Johonson, J. and Reid W.M. (1970): Anticoccidial drugs lesion scoring techniquis in battery and floor Pen experiments with chicken. Exp. Parasitol. 28: 30-36.
- Kim, G.Y.; Kim, S.H.; Hwang, H.; Kim, H.Y. and Park, S.K. (2003): Administration of proteoglycan isolated from *Phellinus linteus* in the prevention of collagen induced arthritis in mice. Boil. Pharm. Bull. 26, 823.
- Kim, S.H.; Songs, S.K.; Kim, B.C.; Lim, J. and Park, EH. (2004): Anti inflammatory and related pharmacological activities of the N-BUOH subfraction of mushroom *Phellinus linteus*. J. Ethnopharmacol. 93, 141.
- Lilleh, J.H.S.; Min, W. and Dalloul, R.A. (2004): Recent progress on cytokine regulation intestinal immune responses to *Eimeria*. Poult. Sci. 83: 611-623.
- Lindeuquist, U.; Teuscher, E. and Nabre (1990): Phytother G. Res. 11, 139. Cited by Hossian M.Z., Akter M.A., Ashraf M.M., and Islam A.K. (2013): Role of oyster mushroom (*Pleurotus ostreatus*) against cecal coccidiosis in cobb 500 broiler chicken. J.Sc., Res. 5(1)185-193.
- Mothana, R.A.; Awadh, N.A.; Jansen, R.; Lindequist, U. and Fitoterpia (2003): Antiviral lanostanoid triterpense from the fungus *Gandoderma pfeifferi*. Biol. Pharm. Bull. 26, 823.
- Naphade, S.T.; Hiware, C.T. and Desarda, S.M. (2010): Studies on the pathological changes during experimental caecal coccidiosis of broiler chicks treated with allopathic amprolium and homoeopathic medicine *Mercurius corrosivus*. Trends Research in Science and Technology 2(1), 49-55.
- Ogbe, A.O.; Uya, I.A.; Ahmed, IL.; Goda; Elisha, D. and Abdu, PA. (2005): A preliminary study on the use of edible mushroom (*Pleurotus ostreatus*) as a source of protein supplement and probiotic in poultry production: implication on human health. Proceeding of

- the 1<sup>st</sup> Nigeria International Poultry Summit (Nips held 20 – 25<sup>th</sup> at Ota, Ogun State Nigeria, PP. 91–95.
- Ogbe, A.O.; Atawod, S.E.; Abdu, P.A. and Itodo, A.E. (2009): Changes in weight gain, faecal oocyst count and packed cell volume of *Eimeria tenella*-infected broilers treated with a wild mushroom (*Ganoderma lucidum*) aqueous extract. J. of south, African Veterinary Association 80(2) 97–102.
- Reshetinkov, S.; Wasser, S.P. and Tan, K.K. (2001): Higher Basidiomycoto as a source of antitumour and immunostimulating polysaccharides. Int. J. Med. Mushroom 3, 361.
- S.A.S (1990): SAS/STAT user's guides. SAS Inst. Inc., Cary, NC. Cited by Doaa A.H., Rania M.E. and H.A. Shalaby (2013): Pathological and biochemical studies on broiler intoxicated with gliotoxin and their control by silicate compounds. Mansoura. Vet. Med. J. (2) 145-160.
- Selegean, M.; Putz, M.V. and Rugea, T. (2009): Effect of polysaccharide extract from the edible mushroom *Pleurotus ostreatus* against infectious bursal disease virus. Int. J. Mol Sci., 10: 3616–3634.
- Soomro, N.M.; Rind, R.; Arijo, A. and Soomro, S.A. (2001): Clinical gross and histopathological studies of coccidial infection in chicken. Int. J. Agri. Biol., 1560: 426-427.
- Sundu, B.; Kumar, A. and Dingle, J.G. (2006): Palm Kernel meal broiler diets: effect on chicken performance and health. World's Poult. Sci. J. 62(2): 316–325.
- Willis, W.L.; Iskhuehmen, O.; Sminor, R.C. and Ohimain, E.I. (2010): Compring the feeding of fungus myceliated grain with other anticoccidial control measures on oocyst extraction of *Eimeria* challenged broilers. Int. J. of Poult. Sci. 9(7): 648-651.
- Willis, W.L.; Wall, D.C.; Iskhuehmen, O.S.; Ibraim, S. and Minor, R.C. (2012): Effect of different mushroom fed to *Eimeria* challenged broilers on reading performance. International J. of Poult. Sci. 11(7): 433-437.
- Yun, C.H.; Lillehoj, H.S. and Lilleuoj, E.P. (2000): Intestinal immuneresponse to coccidiosis. Dev. Comp. Immunol., 24: 303-324.

### دراسات تجريبية على عدوي الكوكسيديا في دجاج التسمين والمعالج بالمستخلص المائي للمشروم

عاده علام عبد الدايم ، دعاء أحمد حسين المطرى ، رحاب رشاد عبد المجيد ، رانيا ابراهيم محمد ابراهيم

Email: [dr.ahmedsob7y@gmail.com](mailto:dr.ahmedsob7y@gmail.com)

أجريت هذه الدراسة للتعرف على تأثير المستخلص المائي للمشروم على الدجاج المصاب بالإيميريا تينبلا ومقارنته بتأثير الامبرول علي معدل الزيادة في الوزن وفي تقليل عدد البويضات التي تخرج من الدجاج وكذلك تحسين التغيرات الباثولوجية في الأعورين. أجريت هذه الدراسة على ١٤٠ من دجاج التسمين عمر ١٤ يوم حيث تم تقسيمها إلى ٧ مجموعات ، يحتوى كل منها علي ٢٠ كتكوت. المجموعة الأولى لم يتم عداها بطفيل الايميريا ولم يتم استخدام أي علاج أو إضافات لها (المجموعة الضابطة) ، المجموعة الثانية تم عداها عند عمر ١٤ يوم بعدد ٥٠٠٠٠٠ حويصلة متجرثمة من الايميريا لكل طائر والمجموعة الثالثة تم عداها كما سبق بنفس الجرعة وفي نفس العمر ثم تم علاجها بإضافة المستخلص المائي للمشروم بجرعة (٢٠٠مجم/ملي) وذلك في اليوم السادس من العدوى ويستمر ذلك لمدة اسبوع متواصل. المجموعة الرابعة يتم عداها كما سبق وفي نفس العمر تم علاجها بالامبرول (٢٠٠مجم/ملي ماء شرب) وذلك في اليوم السادس من العدوى ولمدة اسبوع ، المجموعة السادسة لم يتم عداها بالاييميريا ولكن يتم اعطائها المستخلص المائي للمشروم (٢٠٠مجم/ملي) عند عمر ١٤ يوم ويستمر ذلك يوميا حتى نهاية التجربة. المجموعة السابعة لم يتم عداها بالاييميريا ولكن يتم إضافة الامبرول بجرعة (٢٠٠مجم/ملي ماء الشرب) عند عمر ١٤ يوم ويستمر ذلك حتى نهاية التجربة وقد تم أخذ وزن الطيور أسبوعيا حتى نهاية التجربة. واجراء عدد بويضات الكوكسيديا التي تخرج من البراز في اليوم السابع من العدوى (عند عمر ٢١ يوم) وكذلك بعد اسبوع واسبوعين من العلاج (عند عمر ٢٨ يوم ، ٣٥ يوم) وتم الفحص الباثولوجي في اليوم ٢٨ ، ٣٥ من العمر. وقد أسفرت النتائج عن وجود نقص في وزن الطيور في (المجموعة الثانية) التي تم اصابتها ولم يتم علاجها في كل الأسابيع وكان هناك زيادة معنوية في وزن الطيور التي تم علاجها بالمشروم والامبرول معا (المجموعة الخامسة) وظهر ذلك بوضوح بعد اسبوعين من العلاج (عند عمر ٣٥ يوم) وكان هناك زيادة معنوية في وزن الطيور التي تم علاجها بالمشروم فقط (المجموعة الثالثة) وكذلك هناك زيادة معنوية في وزن الطيور في المجموعة السادسة (التي لم يتم عداها وتم علاجها بالمشروم) وبالتنبيه لعدد الحويصلات فكان أعلى ما يمكن في المجموعة الثانية (التي تم عدوتها ولم يتم علاجها) اما المجموعات التي تم علاجها بالمشروم أو بالامبرول أو بالاثنتين (المجموعة الثالثة والرابعة والخامسة) فكان هناك نقص ملحوظ في عدد البويضات مقارناً بالمجموعة الضابطة. واسفر الفحص الظاهري للاعورين (في المجموعات المصابة) عن وجود انزفة على الجدار الخارجى ووجود زيادة في سمك الجدار ومحتويات الاعورين كانت ما بين اللون البني الى الدم المتجلط، وبالفحص الباثولوجي للمجموعة الثانية فقد وجد نخر وتكسير في الخلايا المحيطة بجدار الاعورين مع وجود مراحل الايميريا بداخلها وتجمع خلايا الليمفوسيت ووجود نزيف وتورم ونخر بالقرب من طبقة العضلات ولكن في المجموعات ٣ ، ٤ ، ٥ والتي تم علاجها بالمشروم أو بالامبرول أو بالمشروم والامبرول معا فقد وجد تحسن ملحوظ في التغيرات الهستوباثولوجية.